



## Effectiveness of SINPV of *Spodoptera litura* (Fab.) (Lepidoptera: Noctuidae) on different host plants

B. S. Ravishankar and M. G. Venkatesha\*

### ABSTRACT

Various bioassay methods were employed to screen SINPV against *S. litura* on different host plants. In the first experiment *S. litura* larvae were reared on different host plants and when they completed second instar, they were transferred to semi-synthetic diet which was treated with different doses of SINPV. The  $LC_{50}$  value against the pest which was reared on different host plants were as follows: cabbage 0.42, cotton 0.61, potato 0.75, groundnut 0.93 and rose 1.28 POB/mm<sup>2</sup>. In the second experiment, larvae of *S. litura* were reared on semi-synthetic diet up to second instar and subsequently they were fed on different host plants, which were treated with SINPV. The  $LC_{50}$  value was highest in rose (1.81) followed by groundnut, potato, cotton and cabbage (0.40 POB/mm<sup>2</sup>). In the third experiment, larvae of *S. litura* were reared on different host plants till they complete second instar and they were screened against SINPV in the laboratory using leaf disc method. The  $LC_{50}$  value was highest in rose followed by groundnut, potato, cotton and cabbage (1.02 POB/mm<sup>2</sup>). In the above three experiments, the  $LC_{50}$  value was lowest in cabbage and was highest in rose.

**Key words:** Host plants, nuclear polyhedrosis virus, *Spodoptera litura*

### INTRODUCTION

*Spodoptera litura* (F.) (Lepidoptera: Noctuidae), commonly known as tobacco caterpillar in India is a major polyphagous pest attacks variety of economically important crops such as cotton, groundnut, rice, tomato, tobacco, citrus, cocoa, potato, rubber, castor, millets, sorghum, maize and many other vegetables (Hill, 1993). Caterpillars of the pest defoliate the crops. The pest occurs in India, Pakistan, Bangladesh, Sri Lanka, S. E. Asia, China, Korea, Japan, Philippines, Indonesia, Australia, Pacific Islands, Hawaii and Fiji (Hill, 1993).

In various species of lepidopteran pest management, increasing failures of chemical pesticides and the problems posed by their indiscriminate use in the field have created a momentum to develop environment friendly methods of pest control. Among the alternatives that are currently available is the use of insect viruses remain the most promising, considering the fact that they can be used in a manner similar to the familiar chemical pesticides. Unlike other natural enemies, insect viruses can be produced and stored and made available to the farmers at short notice due to their longer shelf life. Of the various insect viruses, nucleopolyhedroviruses (NPV) are more successful in pest management (Roberts *et al.*, 1991). *Spodoptera litura* has already developed resistance to several organic pesticides resulting severe crop losses (Singh and Singh, 1998). Fortunately, the pest is highly

susceptible to its NPV and studies have shown that the virus can be used effectively as biopesticide in the field (Jayaraj *et al.*, 1980). *Spodoptera litura* nucleopolyhedrovirus (SINPV) is the most promising control agent and its efficacy has been established successfully against the pest in India (Jayaraj and Rabindra, 1990; Muthuswami *et al.*, 1993). Considering the reliability, suitability and effectiveness of SINPV in terms of economic and ecological reasons, its utilization in pest management has received a great deal of significance.

Although SINPV has been found very effective against *S. litura*, greater amount of variation in the efficacy of SINPV across the host plants hindered its practical utility in the field condition as reported in other lepidopteron pests (Young *et al.*, 1976; Felton *et al.*, 1987; Felton and Duffey, 1990; Keating *et al.*, 1988; Rabindra *et al.*, 1994). Variable efficacy of SINPV and mortality of *S. litura* larvae on different host plants may be related to the secondary metabolites, enzymes, and pH of host plants. Keeping these challenges in mind the preliminary investigations were undertaken to understand the effectiveness of SINPV against *S. litura* on different host plants.

### MATERIALS AND METHODS

Disease-free colonies of *S. litura* were maintained on semi-synthetic diet following the method of Shorey and Hale (1965). In the first experiment, to understand the

effectiveness of SINPV on *S. litura* larvae, which were reared on different host plants and to avoid the direct contact between host plants and SINPV, fresh larvae of *S. litura* from the laboratory culture on semi-synthetic diet were reared on different pot cultured host plants such as cabbage, cotton, groundnut, potato, and rose in the green house. The newly emerged female and male moth from these cultures were paired and caged separately for egg laying. When the eggs hatched the second generation neonate larvae were transferred to the respective host plants and reared up to the end of second instar. When they reached third instar, the larvae of uniform size were selected and introduced singly into vial containing semi-synthetic diet. SINPV was assayed against these larvae on semi-synthetic diet by adopting diet surface contamination method in the laboratory following the earlier procedure (Ignoffo, 1966).

In the second experiment, to understand the effect of host plants on SINPV efficacy by direct contact, lab cultured uniform sized fresh third instar larvae of *S. litura* on semi-synthetic were selected and fed individually with fresh leaf discs (5 mm diameter) of various host plants treated with SINPV for one day in the laboratory. Subsequently, the treated larvae were provided respective fresh leaves (without SINPV) at every 24hrs. In the third experiment, to understand the interaction between host plants (direct contact, host plant secondary metabolites, enzymes, pH) and SINPV, fresh larvae of *S. litura* were reared on different host plants. The newly emerged adults were paired and caged separately for egg laying. When the eggs hatched neonate larvae of second generation were transferred to the respective pot cultured host plants and reared up to the end of second instar. When they reached third instar, the larvae of uniform size were selected and reared on fresh leaf discs of different host plants treated with SINPV. In all the above experiments, to force the larvae to ingest the SINPV treated food, they were kept starved for 2 hr before assaying SINPV as followed earlier (Trang and Chaudhari, 2002). In the above experiments, the larvae reared on respective host plants without treatment of

SINPV were maintained as control. For each treatment, there were three replications with ten larvae per replication. Bioassay of SINPV at different concentrations ( $1 \times 10^6$ ,  $2 \times 10^5$ ,  $4 \times 10^4$ ,  $8 \times 10^3$ ,  $1.6 \times 10^3$ ,  $3.2 \times 10^2$ ) was conducted.  $LC_{50}$  of SINPV was assessed using Probit analysis (Finney, 1952).

## RESULTS AND DISCUSSION

Tobacco caterpillar, *Spodoptera litura* is one of the important agricultural pests. The effect of several control approaches, i.e. chemical control, frequency trembler grid lamps and pheromone traps, the methods based on plant attractiveness and repellency, transgenic plant, and biological control, was studied in order to reduce the population of *S. litura*. Chemical control made a great contribution to suppressing occurrence of *S. litura* and reducing its damage. *Spodoptera litura* larvae reared on different host plants and subsequently fed with semi-synthetic diet treated with SINPV showed significant variations in virulence ( $P < 0.05$ ).  $LC_{50}$  values with respect to different host plants ranged from 0.42 to 1.28 POB per  $mm^2$ . The highest  $LC_{50}$  value was recorded in rose and the lowest was in cabbage (Table 1). Bioassay of SINPV using early third instar larvae of *S. litura* reared on semi-synthetic diet and subsequently fed with SINPV treated leaf discs of various host plants showed significant variations in virulence ( $P < 0.05$ ).  $LC_{50}$  values in different host plants ranged from 0.40 to 1.81 POB per  $mm^2$ . The highest  $LC_{50}$  value was recorded in rose and the lowest was in cabbage (Table 2). Bioassay of SINPV using early third instar larvae of *S. litura* reared on different host plants and subsequently fed with SINPV treated leaf discs of various host plants showed significant variations in virulence ( $P < 0.05$ ).  $LC_{50}$  values in relation to various host plants ranged from 1.02 to 19.50 POB per  $mm^2$ . Again the highest  $LC_{50}$  value was recorded in rose and the lowest was in cabbage (Table 3). In the above experiments, the increasing larval mortality was noticed from fourth to tenth day after treatment. Moreover, the larval mortality was also increased with increasing concentrations of SINPV irrespective of the host plants.

**Table 1.** *Spodoptera litura* reared on different host plants and assayed against SINPV on semi-synthetic diet.

Host plants	$\chi^2/(n-2)$	Slope $\cdot b' \pm S.E$	$LC_{50}$ (POB/ $mm^2$ )	Fiducial limits		$LC_{99}$ (POB/ $mm^2$ )
				Lower	Upper	
Cabbage	0.51	$0.86 \pm 0.10$	0.42	0.22	0.76	201.65
Cotton	0.75	$0.92 \pm 0.11$	0.61	0.34	1.09	199.64
Potato	1.80	$0.73 \pm 0.09$	0.75	0.38	1.49	1124.63
Groundnut	0.15	$0.72 \pm 0.09$	0.93	0.47	1.87	1459.69
Rose	0.48	$0.66 \pm 0.09$	1.28	0.62	2.80	4028.39

\*All lines are significantly a good fit ( $p < 0.05$ )

**Table 2.** *Spodoptera litura* reared on semi-synthetic diet and assayed against SINPV on different host plants.

Host plants	$\chi^2/(n-2)$	Slope 'b' ± S.E	LC <sub>50</sub> (POB/mm <sup>2</sup> )	Fiducial limits		LC <sub>99</sub> (POB/mm <sup>2</sup> )
				Lower	Upper	
Cabbage	0.40	0.70 ± 0.09	0.40	0.19	0.81	848.87
Cotton	0.47	0.71 ± 0.09	0.81	0.40	1.64	1501.96
Potato	1.69	0.55 ± 0.08	0.94	0.40	2.31	14136.66
Groundnut	0.22	0.70 ± 0.09	1.36	0.68	2.86	2784.34
Rose	0.84	0.63 ± 0.09	1.81	0.86	4.25	8136.75

\*All lines are significantly a good fit (p<0.05)

**Table 3.** Effectiveness of SINPV against *S. litura* on different host plants.

Host plants	$\chi^2/(n-2)$	Slope 'b' ± S.E	LC <sub>50</sub> (POB/mm <sup>2</sup> )	Fiducial limits		LC <sub>99</sub> (POB/mm <sup>2</sup> )
				Lower	Upper	
Cabbage	0.84	0.60 ± 0.09	1.02	0.47	2.33	6768.36
Cotton	0.44	0.62 ± 0.09	2.27	1.07	5.56	11495.64
Potato	0.14	0.48 ± 0.08	4.48	1.72	17.60	255101.49
Groundnut	0.88	0.46 ± 0.08	8.14	2.83	44.60	865931.08
Rose	0.23	0.35 ± 0.08	19.50	4.58	443.06	61947027.97

\*All lines are significantly a good fit (p<0.05)

The results of all the experiments conducted to know the influence of host plants on the effectiveness of SINPV against *S. litura* larvae revealed that there was a significant variation in the LC<sub>50</sub> values among different host plants and it was lowest in cabbage and highest in rose. Similarly, LC<sub>50</sub> values of SINPV screened against *S. litura* varied in different host plants (Kulkarni, 1997). It is known that the leaf exudates from glandular hairs of the cotton plants inactivate the NPV of *Heliothis* spp. (Falcon, 1971 and Young and Yearin, 1974). Further, it is reported that plants mediate interactions between insect and its pathogens, which increase or decrease the impact of the insect pathogen (Rabindra *et al.*, 1994; Meade *et al.*, 1995). From the present experiments it could be made out that different host plants influence the effectiveness of SINPV by direct or indirect means. Further investigations on the leaf pH and secondary metabolites of different host plants may provide what are the factors interfere with the virulence of SINPV. Eight of baculoviruses genes help the virus overcome insect defenses (Suzanne M. Thiem, 2009).

#### ACKNOWLEDGEMENT

We are highly grateful to Dr. K.P. Jayanth, Vice President, Bio-control Research Laboratory (BCRL), Pest Control of India Private Limited (PCI), Bangalore for providing the host insect, virus and laboratory facilities for conducting the experiments.

#### REFERENCES

Falcon, L. A. 1971. Microbial control as a tool in integrated

control programs. **In:** *Biological Control*, (Huffaker, C.B.ed.). Plenum Press, New York. 346-364 **PP**.

Felton, G. N. and Duffey, S. S. 1990. Inactivation of baculovirus by quinines formed in insect damaged plant tissues. *Journal of Chemical Ecology*, **16**: 1221-1236.

Felton, G. W., Duffey, S. S., Vail, P. V., Kaya, H. K. and Manning, J. 1987. Interaction of nuclear polyhedrosis virus with catechols: Potential incompatibility for host plant resistance against noctuid larvae. *Journal of Chemical Ecology*, **13**: 947-957.

Finney, D. J. 1952. Probit analysis. Cambridge University Press, London, UK, 151 **P**.

Hill, D. S. 1993. *Agricultural insect pests of the tropics and their control*. Cambridge University Press: Cambridge, London.

Ignoffo, C. M. 1966. Effects of temperature on mortality of *Heliothis zea* larvae exposed to sublethal doses of nuclear polyhedrosis virus. *Journal of Invertebrate Pathology*, **8**: 290-292.

Jayaraj, S. and Rabindra, R. J. 1990. Microbial control and integrated pest management. Proceedings of indo USSR joint workshop on problems and potentials of Biocontrol of pests and Diseases, Bangalore, 263-387 **PP**.

Jayaraj, S., Santharam, S., Narayan, K., Soundararajan, K. and Balagurunathan, R. 1980. Effectiveness of the nuclear polyhedral virus against field populations of the tobacco caterpillar, *Spodoptera litura* on cotton. *The Andhra Agricultural Journal*, **27**:26-29.

- Keating, S. T., Hunter, H. D. and Schultz, J. C. 1988. Relationship between susceptibility of gypsy moth larvae (Lepidoptera: Lymantridae) to a baculovirus and host plant constituents. *Environmental Entomology*, **17**: 952-958.
- Kulkarni, G. G. 1997. Influence of host plants in the multiplication and persistence of *Spodoptera litura* Fab. NPV. M.Sc. Thesis University of Agricultural Sciences, Dharwad.
- Meade T., Felton, G. W. and Young. S. Y. 1995. Host plant modification of susceptibility to the *Heliothis* nuclear polyhedrosis virus, In: proceedings of the 1995 Cotton Research Meeting, held at Mississippi county Community College, Blytheville. 9 February, **172**: 63-65.
- Muthuswami, M., Rabindra, R. J. and Jayaraj, S. 1993. Use of baculovirus mixture for the control of *Helicoverpa armigera* and *Spodoptera litura* on Groundnut. *Journal of Biological Control*, **7**: 105-108.
- Rabindra, R. J., Muthusami, M. and Jayaraj, S. 1994. Influence of host plant surface environment on the virulence of nuclear polyhedrosis virus against *Helicoverpa armigera* (Hbn) (Lepidoptera: Noctuidac) larvae. *Journal of Applied Entomology*, **118**: 453-460.
- Roberts, D. W., Fuxa, J. R., Gaugler, R., Goettel, M, Jaques R. and Maddox. J. 1991. Use of pathogens in insect control, **In**: (Pimentel, D. ed.). *CRC Handbook of Pest Management in Agriculture*, second edition, Boca Raton, Fla, CRC Press. **2**: 243-278 PP.
- Shorey, H. H. and Hale, R. L. 1965. Mass rearing of larvae of nine noctuid species on a simple artificial medium. *Journal of Economic Entomology*, **58**: 522-524.
- Singh, D. P. and Singh, J. P. 1998. Relative susceptibility and development of resistance in *Spodoptera litura* larvae against some pyrethroids and non-pyrethroid insecticides. *Indian Journal of Entomology*, **60**: 177-180.
- Suzanne M. Thiem, 2009. Baculovirus genes affecting host function. *In vitro Cellular and Developmental Biology - Animal*, **45**: (3 & 4): 111-126.
- Trang, T. T. K. and Chaudhari, S. 2002. Bioassay of nuclear polyhedrovirus (npv) and in combination with insecticide on *Spodoptera litura* (Fab), *Omonrice*, **10**: 45-53.
- Young, S. Y. and Yearian, W. C. 1974. Persistence of heliothis NPV on foliage of cotton, soybean and tomato, *Environmental Entomology*, **3**: 253-255.
- Young, S. Y., Yearian, W. C. and Kim, K. S. 1976. Effect of dew from cotton and soybean foliage on activity of *Heliothis nuclear polyhedrosis virus*. *Journal of Invertebrate Pathology*, **29**: 105-111.

**B. S. Ravishankar and M. G. Venkatesha\***

Department of Zoology, Bangalore University, Jnana Bharathi, Bangalore - 560 056, Karnataka, India, \*E-mail: venkatatmelally@gmail.com