



## ***In vitro* inhibitory effect of fungicides and botanicals on mycelial growth and spore germination of *Fusarium oxysporum***

Taskeen-Un- Nisa, A. H. Wani, Mohd Yaqub Bhat, S.A. Pala and R. A. Mir

### **ABSTRACT**

Carbendazim, hexaconzole, bitertanol, myclobutanil, mancozeb, captan and zineb and extracts of *Allium sativum*, *Allium cepa* and *Mentha arvensis* were evaluated for their effect on the inhibition of mycelial growth and spore germination of *Fusarium oxysporum*. Maximum inhibition in mycelial growth was observed in the hexaconazole at 1000 ppm followed by other fungicides at the same concentration. In case of botanicals, inhibition in spore germination was highest at concentration 'S'. It was followed by S/2, S/10, and S/100 concentrations of plant extracts as compared to control which showed least inhibition in spore germination.

**Key words:** *In vitro*, botanicals, fungicides, *Fusarium oxysporum*, mycelial growth, spore germination.

### **INTRODUCTION**

Many fungi have been identified by various workers as causal organism of fungal rot diseases in all parts of the world. The principal rot diseases on tomato and other vegetables with varying intensities include early blight or *Alternaria* rot caused by *A. solani* (Ell. and Mart.) Jones and Grout, *A. tenuis* (Nees) Syn. and *A. alternata* (Fr.) Kessel, Late blight or *Phytophthora* rot caused by *Phytophthora infestans* (Mont.) Debary. *Alternaria* rot has been considered as the most common disease of tomato and other plants and causes heavy losses in quality of the fruits, thus rendering large quantity of tomato fruits unfit for consumption (Barker and Fauchs, 1980.; Hassan, 1996; Singh *et al.*, 1997). Chemical control measures have been tested and found effective in the control of diseases (Ogundana and Denis, 1981; Plumbley, 1985). Certain protective fungicides although hazardous to environment are still used for the control of fungal diseases (Nwankiti *et al.*, 1990; Vaish and Sinha, 2003). Likewise, use of pesticides of plant origin have been suggested by some workers as alternative to synthetic chemicals in order to counter the potential hazardous effect on the environment associated with the use of synthetic chemicals (Amadioha and Obi, 1999; Ejechi and Ilondi, 1999; Singh, *et al.*, 1997; Amadioha, 2000). Therefore, in the present investigation, inhibition of mycelial growth and spore germination of important post harvest fungus, *Fusarium oxysporum*, exposed to different concentrations of some fungicides and plant extracts of some plants were studied. The aim of the present study was to compare the effect of some selected fungicides and plant extract on *Fusarium oxysporum* mycelial growth and spore

germination *in-vitro* and identify the concentration of plant extract that have fungicidal properties.

### **MATERIALS AND METHODS**

Systemic and non-systemic fungicides *viz.*, carbendazim, myclobutanil, bitertanol, hexaconazole, mancozeb, captan and zineb were evaluated for their efficacy on mycelial growth of *Fusarium oxysporum* by food poisoning technique (Falck, 1907; Grower and Moore, 1962). Appropriate quantity of each fungicide was separately dispensed in molten sterilized PDA medium to make desired concentrations for each fungicide. The mycelial discs of 5 mm diameter, taken from ten days-old culture of the fungal pathogens were aseptically placed in the center of solidified poisoned PDA. Five replications were maintained for each concentration. The Petri-plates were incubated at  $24 \pm 2$  °C and observations on the mycelial growth of test fungus were recorded after seven days of incubation. The growth of test fungus on non-poisoned PDA served as a control. The percent inhibition in growth due to various fungicidal treatments at different concentrations was computed as follows: Mycelial growth inhibition (%) =  $[(dc - dt) / dc] \times 100(\%)$

Where dc = average diameter of fungal colony in control, and dt = average diameter of fungal colony in treatment group

In the present study different concentrations of aqueous extracts of plant leaves and bulbs of mint, *Mentha arvensis* L. onion, *Allium cepa* L. and garlic were evaluated for their effect on the spore germination of *F. oxysporum*. For the preparation of different concentrations of plant extracts, 200g each of leaves and bulbs were washed with sterilized distilled water,

grinded in Mortor and pestle using 200 ml of sterilized distilled water (Bhat and Sivaprakasan, 1994). The material was homogenized for 5 minutes and filtered through double layered muslin cloth followed by Whattman’s filter paper No. 1. The filtrate was then centrifuged at 5000 rpm for 10 minutes and considered as standard solutions (S). The solution diluted 2 times (S/2), ten times (S/10), and hundred times (S/100) with sterilized distilled water, and were used to study the spore germination of *F. oxysporum*.

Spore suspension of each isolate of fungus containing at least 20-30 spores per microscopic field was prepared from ten day-old fungal culture. One drop of about 0.1ml of spore suspension was placed in a cavity glass slide containing a drop (about 0.1ml) of different concentration of plant extract. These slides were kept in moist chamber prepared by putting two folds of filter paper in both sides of Petri-plates. These Petri plates were incubated at 24±2 °C for 24 hr. Each treatment was replicated five times. The percent spore germination was recorded using formula given by Kiraly *et al.* (1974).

$$\text{Percent spore germination} = \frac{\text{No. of spores germinated}}{\text{Total no. of spores examined}} \times 100$$

The data collected during these investigations were subjected to appropriate statistical analysis using Minitab software. The data wherever needed was subjected to appropriate transformation as suggested by Gomez and Gomez (1984) before statistical analysis. The method given by Panse and Sukhatme (1978) was also used for statistical analysis of the data.

**RESULTS AND DISCUSSION**

**Fungicides on the mycelial growth**

It was revealed from the results (Table 1) that all systemic fungicides at different concentrations significantly inhibit the mucelial growth of *F. oxysporum*. However, the hexaconozole at highest concentration (1000 ppm) caused highest reduction of mycelail growth (8.80 mm) followed by carbendazim (9.40 mm), bitertanol (18.60 mm) and myclobutanil (20 mm) at the same concentration. It was also observed from the study that amongst the non-systemic fungicides, mancozeb was found most effective (14.20mm) in reducing mycelia growth of the fungi followed by captan (20.00 mm) and zineb (22.00 mm). Similar finding were reported by Daradhiyar (1980); Sommer (1982); Kalra and Sokhi (1985); Singh *et al.* (1997); Patel *et al.* (2005) and Banyal *et al.* (2008) to other fungi. However, the fungicides have been shown to completely inhibit the mycelial growth of *Fusarium oxysporum* in Richard medium (Khan *et al.*, 1997; Sharma, 2006).

**Plant extracts on the spore germination**

It was revealed from the results (Table 2) that different concentrations of plant extracts caused significant inhibition in the spore germination. However, the maximum inhibition in the spore germination was found at highest concentration ‘S’ followed by S/2, S/10, and S/100 as compared to control which showed least inhibition in spore germination. The extract of *A. sativum* at highest concentration (‘S’) was found to be most effective in reducing the spore germination followed by highest concentration (S) of extract of *A. cepa* and *M. arvensis*. The inhibition in spore germination varies from 43.94% to 9.7% in different concentrations of *A. sativum*. In different concentrations of extract of *Allium cepa*, the inhibition in spore germination ranges from 60.51% to 17.58% whereas

**Table1.** Effects of different concentrations of fungicides on the mycelia growth of *Fusarium oxysporum*

Concentration Treatment	Mycelial growth (mm)*				
	125ppm	250ppm	500ppm	1000ppm	Control
Systemic fungicide					
Carbendazim	24.00	19.40	14.80	9.40	69.40
Bitertanol	33.00	28.60	21.60	16.80	71.60
Myclobutanil	36.00	32.40	27.40	18.60	71.80
Hexaconozole	23.60	16.80	12.20	8.80	67.80
Non-systemic fungicide	500ppm	1000ppm	1500ppm	2000ppm	Control
Mancozeb	31.40	21.60	17.40	14.20	65.80
Zineb	42.00	35.20	27.00	22.00	70.20
Captan	34.60	29.40	25.00	20.00	70.20

**Table 2.** Effect of plant extracts on the spore germination of *Fusarium oxysporum*.

Concentration Treatment	Spore germination (%)*				
	Control	S/100	S/10	S/2	S
<i>Allium sativum</i>	90.50 (72.06)	43.94 (41.52)	32.60 (34.82)	21.29 (27.48)	9.7 (18.20)
<i>Allium cepa</i>	91.90 (73.47)	60.51 (51.07)	51.93 (46.26)	41.26 (39.97)	17.58 (24.79)
<i>Mentha arvensis</i>	92.57 (74.19)	69.39 (56.41)	58.61 (49.96)	39.26 (38.80)	20.44 (26.88)

Value in parentheses indicates

inhibition of spore germination ranges from 69.39% to 20.44% in different concentration of extract of *Mentha arvensis* respectively as compared to untreated control which showed least inhibition in spore germination.

Previously 31 plants belonging to Asteraceae family (Rai and Acharya, 1999) were tested against the cowpea wilt pathogen, *Fusarium oxysporum* f. sp. *ciceris*. Ozer *et al.* (2003) evaluated the pectolytic impact of *Allium cepa* "Akugun 12" against two *Fusarium* isolates FOC6 and FOC8. In addition *A. sativum* was known to act as anti-fungal activity (Sahavaraj *et al.*, 2006). Similar results were found by Misra and Dixit (1976) and Bowers and Locke (2000) using *Allium sativum* against eighteen different fungi including *Fusarium* spp.. Jacob and Siva Prakashan (1994) and Arya *et al.* (1995) studied the antifungal activity of the extracts of various plant species against *Fusarium pallidoroseum* and reported inhibitory effect of extracts of garlic bulbs and *Bignonia* leaves on the mycelial growth of *Fusarium pallidoroseum*. Karade and Sawant (1999), Datar (1999) and Anwar and Khan (2001) observed the same results with the plant extracts of other plants. Our findings are also in agreement with those of Bansal and Gupta. (2000), Bashir (2001) and Bhat (2002). Tejada Meneses *et al.* (2002) reported antifungal activity of some wild plants against *Penicillium* sp.

In conclusion it was revealed from the study that both fungicides and extracts of plant origin caused inhibition in mycelial growth and spore germination of *Fusarium oxysporum*. However, plant extracts utilized above in this study for *in-vitro* inhibition of spore germination can prove better substitute under field conditions to the management of pathogenic fungi with fungicides which are hazardous to the environment.

## REFERENCES

- Amadioha, A.C. 2000. Controlling rice blast *in vitro* and *in vivo* with extracts of *Azadirachta indica*. *Crop Protection*, **19**: 287-290.
- Amadioha, A. C. and Obi, V. I. 1999. Control of Anthracnose disease of cowpea by *Cymbopogon cunitus* and *Ocimum gratissimum*. *Acta Phytopathology Entomology Hungary*, **85**: 89.
- Anwar, A. and Khan, F.U. 2001. Effect of aqueous leaf extracts of medicinal plant on the growth of rhizospheric fungi of tomato cv. Pusa Ruby *in vitro*. *SKUAST Journal of Research*, **3**: 60 - 63.
- Arya, A., Chauhan, R. and Arya, C. 1995. Inhibition of growth of 200 pathogenic fungi by garlic extract. *Mycologia*, **67**: 882 - 885.
- Bhat, Z.A. 2002. Comparative efficacy of bio-control agents, Botanical extracts and fungicide in the management of chickpea wilt caused by *Fusarium oxysporum*. M. Sc. (Ag.) thesis, Allahabad Agriculture Institute (Deemed University). Allahabad-211007, (U.P) India. 65pp.
- Bhat, N.M. and Sivaprakashan, K. 1994. Antifungal activity of some plant extracts. In: Crop innovation techniques and management (Sivaprakashan, K. and Seetharaman, I. eds.). Kalyani publishers, New Delhi, India, 335-339 **PP**.
- Bansal, R.K. and Gupta, R.K. 2000. Evaluation of plant extracts against *Fusarium oxysporum*. *Indian Phytopathology*, **35**: 107-108.
- Banyal, D.K. Mankotia, V. and Sugha, S.K. 2008. Integrated management of tomato collar rot caused by *Sclerotium rolfsii*. *Journal of Mycology and Plant Pathology*, **38**(2): 165-167.
- Barker, G. R. and Fauchs, Y. 1980. Research on postharvest diseases of tomato. A survey of the organisms causing rot of stored tomato fruits. *Preliminary report, volcane centre, Bet-Dagan*, **782**: 1-12.
- Bashir, S. 2001. Evaluation of some medicinal plant extracts against *Fusarium oxysporum* f. sp. and *Alternaria* sp. M.Sc (Ag) Thesis, Allahabad Agriculture Institute (Deemed University), Allahabad, U.P, India, 65 **P**.
- Bowers, J.H. and Locke, 2000. Effect of botanical extracts on the population density of *Fusarium oxysporum* in soil and control of *Fusarium* wilt in the green house. *Plant Disease*, **3**: 300-305.
- Daradhiyar, P. K. 1980. Studies on some post harvest disease of tomato and their control. *Journal of Indian Botanical Society*, **59**(3): 230-233.
- Datar, V.V. 1999. Bioefficacy of plant extracts against *Macrophomona phaseolina* (Tassi) Goid, the incitant to charcoal- rot of sorghum. *Journal of Mycology and Plant Pathology*, **29**: 251-253.

- Douglas, B. 1922. A new *Alternaria* spot of tomatoes in California. *Phytopathology*, **12**: 146-148.
- Ejechi, B.O. and Iiondi M.E. 1999. Control of yam tuber (*Dioscorea rotundata*) rot agent *Sclerotium rolfsii* with Camwood (*Baphida nitida* Lodd) sawdust extract. *African Journal of Root Tuber Crops*, **3**(2): 13-15.
- Falck, R. 1907. Wachstumsgesetze, wachstum Laktorehnund temperature wertder holzersterenden. *Myceture*, **32**: 38-39.
- Gomez, K. A. and Gomez, A. A. 1984. Statistical producers for agricultural research. Jhon Wiley and Sons Inc., 680 **PP**.
- Hassan, H.A.H. 1996. *Alternaria* mycotoxins in Black rot lesion of tomato fruits: Conditions and regulations of their productions. *Acta Immunologica Hungarica*, **43**(2-3): 125-133.
- Jacob, C.K. and Sivaprakasam, K. 1994. Evaluation of some plant extracts and antogonsists for the control of pre-emergence damping-off of brinjal (*Solanum melongena* L.) In: Crop Disease-innovative Techniques and management (Sivaprakaasam, K. ed.). Kalayni Publisher, New Dehli, 289-294 **PP**.
- Kalra, J. S. and Sohi, H. S. 1985. Studies on post-harvest rot of tomato fruits. Control of *Alternaria* fruit rot. *Indian Journal of Mycology and Plant Pathology*, **15**(3): 256-260
- Karade, V. M. and Sawant, D.M. 1999. Effect of some plant extract on the spore germination of *Alternaria alternata*. *Plant Disease Research*, **14**: 75-77
- Khan, M. A., Ahmad, M. and Saeed, M. A. 1995. Evaluation of fungicides on the growth of *Alternaria alternata* in-vitro and the control of the post harvest tomato fruit rot. *Pakistan Journal of Phytopathology*, **7**(2): 166-168.
- Kiraly, Z., Klement, S.J., Voros and Solymosy, K. 1974. Methods in plant pathology with special reference to breeding for resistance to breeding for resistance. Elsevier scientific publishing company, New York, 212 **PP**.
- Misra, S.B. and Dixit, S.N. 1976. Fungicidal spectrum of the leaf extract of *Allium sativum*. *Indian Phytopathology*, **29**: 448-449.
- Nwankiti, A.O., Kalu, A.D. and Ene, L.S.O. 1990. Seed yam production by minisett technique. Varietals responses to curing treatment as alternative to chemical seed dressing. *Nigerian Journal of Plant Protection*, **13**: 1-5.
- Ogundana, S.K. and Denis, C. 1981. Assessment of fungicides for prevention of storage rot of yams. *Pesticide Science*, **11**: 491-494.
- Ozer, N., Koyca, D., Chiloni, G., Pizzuolo, P.H., Coskuntuna A. and Magro, P. 2003. Pectolytic isoenzymes by *Fusarium oxysporum* f sp. *cepae* and antifungal compounds in onion cultivars as a response to pathogen infection. *Canadian Journal of Plant Pathology*, **25**: 249-257.
- Panse, V.G. and Sukhatme, P. V. 1978. *Statistical Methods for Agricultural Workers*. Indian Council of Agricultural Research, New Delhi, 358 **P**.
- Patel, N. A., Dange S.R.S. and Patel, S.I. 2005. Efficacy of chemicals in controlling fruit rot of tomato caused by *Alternaria* tomato. *Indian Journal of Agricultural Research*, **39**: 1.
- Plumbley, R.A. 1985. Benomyle tolerance in strain of *Penicillium scleroteginum* infecting yams and use of imazalid as a means of control. *Tropical Agriculture Trinidad*, **61**: 182-185.
- Rai, M.K. and Acharya, D. 1999. Screening of some Asteraceous plants for antimycotic activity. *Compositae Newsletter*, **34**: 37-43.
- Sahayaraj, K., Namasivayam, S.K.R. and Borgio, J. A. F. 2006. Influence of three plant extracts on *Fusarium oxysporum* f. sp. *ciceria* mycelium growth. *Journal of Plant Protection Research*, **46** (4): 335 – 338.
- Sharma, R.L. 2006. Efficacy of fungicide impregnated paper liners against storage rot of tomato fruit. *Journal of Mycology and Plant Pathology*, **26** (2): 310-311.
- Singh, S.N., Yadav, B.P., Sinha, S.K and Ojha, K.L. (1997). Efficacy of plant extract in inhibition of radial growth of *Colletotrichum capsici*. *Journal of Applied biology*, **51**:180-183.
- Sokhi, S.S. and Sohi, H.S. 1972. *Rhizoctonia* fruit rot of tomato. *Indian journal of Mycology and Plant Pathology*. **2**: 191-192.
- Sommer, N.F. 1982. Post harvest handling practices and post harvest diseases of fruits. *Plant Disease*, **66**: 357-364.
- Tequida meneses, M., Cortez R. M., Rosas burgos, E.C. and Corrales, M.C. 2002. Effect of alcoholic extracts of wild plants on the inhibition of growth of *Aspergillus niger*, *Penicillium chrsogenum*, *Pencillium expansum* and *Fusarium poae* molds. *Revista Ibetroamericana Micologia*, **19**: 84-88.
- Vaish, D.K. and Sinha, A.P. 2003. Determination of tolerance in *Rhizocotnia solani*, *Trichoderma virens* and *Trichoderma* sp. (isolate 20) to systemic fungicides. *Indian journal of Plant Pathol.* **21**(1-2):48-50

---

**Taskeen-Un- Nisa, A. H. Wani, Mohd Yaqub Bhat\*, S.A. Pala and R. A. Mir**

Section of Plant Pathology and Mycology, P.G. Department of Botany, University of Kashmir, Srinagar - 190006, India, \*Corresponding author E-mail: myaqub35@gmail.com