JBiopest 15(2):129-135(2022)

The sequence between mitochondrial nad1-nad2 genes are potential molecular markers of harpactorinae predators (Heteroptera: Reduviidae)

Sherlin John, J^{1,2,}., Selvaraj, P^{2*}., Pushpanathan, T²., Ravichandran, B³., Sayed AbdulAzeez⁴ and Francis Borgio, J⁴.

ABSTRACT

Assassin bugs are terrestrial predators belonging to the family Reduviidae. Among the 24 subfamilies, harparctorinae is predominant and extensively investigated for biocontrol applications compared to the others subfamilies in Reduviidae. Being natural enemies of phytophagous insects. understanding the phylogeny of these predatory bugs can precise the selection of candidates to employ in insect pest management. In addition to morphological systematics, complete mitochondrial genome sequences provide great insights into the phylogeny for resolving evolutionary complexity. Complete mitochondrial genomes of four potential predatory harpactorinae and one outgroup triatominae were retrieved from NCBI GenBank database. Comparative analysis of the five mitogenomes and the nucleotide sequence between nad1 and nad2 genes were selected as the best option to distinguish. The nucleotide sequence between nad1nad2 are found to be biased towards A and T similar to their respective complete mitogenomes. Tajima's test of neutrality suggest that the evolutionary selection at nad1-nad2 was parallel to the complete mitogenome and showed positive and significant (p > 0.1) with high nucleotide diversity. Unequal evolutionary rate at nad1-nad2 between lineages observed in Tajima's relative rate test and proved the nucleotide sequences of *nad1-nad2* between species are highly variable. Comparing the phylogenetic trees generated using the complete mitogenomes and nad1-nad2 genes uncovered the correlation between the trees and having identical branches with varying bootstrap values. Conventionally the highly conserved protein-coding cox1 gene is used for molecular taxonomy whereas this study provides an additional and/or a possible alternative molecular marker for genetic comparative test (the nucleotide sequence between nad1-nad2) to understand the systematics and phylogeny of Reduviidae. The significant nucleotide diversity, high genetic distance and less genetic similarity of the sequence between nad1-nad2 genes among the species studied, Agriosphodrus dohrni, Rhynocoris fuscipes, Scipinia horrida, and Velinus nodipes undoubtedly propose the possible utilization of nad1-nad2 region as distinguishable molecular

Keywords: Reduviidae, Biocontrol agent, Mitochondrial genome, Molecular markers, Phylogeny

MS History: 20.10.2022(Received)-01.11.2022(Revised)- 22.11.2022 (Accepted)

Citation: Sherlin John, J., Selvaraj, P., Pushpanathan, T., Ravichandran, B., Sayed AbdulAzeez and Francis Borgio, J. 2022. The sequence between mitochondrial nad1-nad2 genes are potential molecular markers of harpactorinae predators (Heteroptera: Reduviidae). *Journal of Biopesticides*, **15**(2): 129-135.

DOI: 10.57182/jbiopestic.15.2.129-135

INTRODUCTION

Assassin bugs are terrestrial predatory heteropterans belonging to the family Reduviidae.

There are about 7000 species including subspecies in 24 sub-familes. With the exception of hematophagous triatominae all the other

subfamilies are predatory. Harpactorinae is the largest subfamily consisting of morphologically diverse species about 2,800 in number across 300 genera (Ambrose, 1999; Weirauch et al., 2014). They are generalist predators feeding on multiple phytophagous insects through different orders viz., Coleoptera, Hemiptera, Hymenoptera, Dictyoptera, Lepidoptera, Orthoptera Arachnids. Being natural enemies of many insects, their diurnal behaviour and plant-dwelling habit, the harpactorinae assassin bugs are proved to be biocontrol agents and potential augmented successfully in control of many pests more than the other subfamilies of Reduviidae (Ambrose, 1995; Sahayaraj, 2007 and 2014; Schuh and Weirauch, 2020). For example, Agriosphodrus dohrni employed in the control of forest pest forest pest Hyphantria cunea (Du et al., 2018); Rhynocoris fuscipes against groundnut pests such as Aphis craccivora, Spodoptera litura and Mylloceros sp. (Sahayaraj et al., 2002); Scipinia horrida on psyllid *Heteropsyila cubana* (Barrion et al., 1987) and Velinus nodipes versus Linaeideu aenea (Takeno, 1998). However, research around the world for the utilization of assassin bugs as biocontrol agents is lag (Ambrose and Kumar, 2016). Exploration into any group of living organisms starts with proper identification and classification as it is fundamental to the study of life science (Quicke, 1993). Sole dependance on classical taxonomy is time consuming and is not valued by the modern scientific community. Application of molecular technology systematic studies solve the evolutionary complexities in cryptic groups (Yi et al., 2022). systematics Molecular relies heavily mitochondrial genomes due to their conserved nature across species. In addition to morphological systematics, complete mitochondrial genome sequences resolve evolutionary complexities (Kjer, 2016). Typical invertebrates including hemipteran mitogenomes feature 13 protein-coding genes, 22 transfer RNA genes and 2 ribosomal RNA subunit genes (Cameron, 2014i and 2014ii; Wang et al., 2015). Conserved protein-coding genes such as cox1 are used in molecular identification i.e., DNA

barcoding (Jung, 2010). This study probes the nucleotide sequences between the protein-coding genes and their chances of being utilized as species specific molecular markers. Preliminary investigations with Triatominae suggest the highly variable nucleotide sequences between *nad1-nad2* (Selvaraj *et al.*, 2022) were useful and that the phylogenetic analysis in this study can validate its significance.

MATERIALS AND METHODS

Complete mitochondrial genomes of biocontrol subfamily from the Harpactorinae (Heteroptera: Reduviidae) catalogued in the NCBI Nucleotide database (https://www.ncbi.nlm.nih.gov/nuccore) were downloaded. The selected species with their GenBank accession number are A-Agrisphodrus B-Rhynocoris fuscipes (NC015842), dohrni (MZ440304), C-Scipinia horrida (NC037744), D-Velinus nodipes (NC037738) and outgroup E-Rhodnius prolixus (NC050328). Circular genome map was created using CGview (https://proksee.ca/). Dataset FASTA files were generated for analysis using Unipro **UGENE** software (http://ugene.net/). Multiple sequence alignment was performed using MAFFT 7 online server (https://mafft.cbrc.jp/). Tajima's test for neutrality and relative rate was computed in MEGA 11 (Tamura et al., 2021) with codon positions 1st+2nd+3rd+Noncoding. included were positions containing gaps and missing data were eliminated (complete deletion option). Trees were constructed using neighbourhood-joining method, substitution model Jukes-Cantor and 200 Bootstrap resampling. The constructed trees were processed in **ITOL** web server (https://itol.embl.de/). The results were presented as tables and graphs using Microsoft Office Excel 16.

RESULTS

Nucleotide composition

The nucleotide sequence between *nad1-nad2* features five transfer RNAs (Leucine-1, Valine, Isoleucine, Methionine and Glutamine), 2 ribosomal RNA subunits and a control region (Figure 1).

131

Table 1. Nucleotide composition of the selected complete mitogenome sequence and the sequence between *nad1-nad2* of harpactorinae biocontrol agents.

Species	Length (bp)		A%		T(U)%		G%		С%	
	Complete	nad1- nad2	Complete	nad1- nad2	Complete	nad1- nad2	Complete	nad1- nad2	Complete	nad1- nad2
Agriosphodrus dohrni	16470	4039	38.99	39.47	33.22	33.97	12.19	10.47	15.60	16.09
Rhynocoris fuscipes	15542	3234	40.94	40.29	31.08	32.53	11.50	10.14	16.48	17.04
Scipinia horrida	15660	3376	40.78	39.90	32.48	34.06	11.35	10.28	15.38	15.76
Velinus nodipes	15904	3458	41.13	41.82	32.11	32.94	11.50	9.86	15.26	15.38
Average	15894	3527	40.46	40.37	32.22	33.38	11.63	10.19	15.68	16.07
Rhodnius prolixus*	15789	3524	41.03	40.87	28.57	30.77	11.05	9.82	19.35	18.54

(* Outgroup)

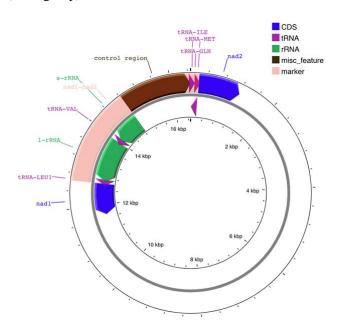


Figure 1. Representative circular map of the harpactorinae mitogenome showing the features present between the genes *nad1-nad2*.

Agriosphodrus dohrni has the largest (16,470 bp) mitgenome among the reported mitogenome of harpactorinae. The length of the complete mitogenome sequences of the harpactorinae biocontrol agents taken for the study vary from 15,542 bp (*Rhynocoris fuscipes*) to 16,670 bp (*Agriosphodrus dohrni*) with an average 15,894

bp. Whereas, the length of the sequence between the genes *nad1-nad2* range from 3,324 bp (Rhynocoris fuscipes) to 4,039 bp (Agriosphodrus Base dohrni). composition of complete mitogenome revealed that the percentage of adenosine (A 40.46 %) is predominant over the other nucleotides (T 32.22T% > C 15.68% > G 11.63%). Similar characteristics on the composition of nucleotides in the sequence between nad1-nad2 was observed (A 40.37% > T $33.38\% > C \ 16.07\% > G \ 10.19\%$) (Table 1). The difference among nucleotide composition of the selected four species of harpactorinae on the complete mitogenome and nad1-nad2 revealed their genetic conservation.

Test of neutrality for evolutionary selection

Test of neutrality analysis of nad1-nad2 genes of 5 nucleotide sequences (4 Harpactorinae and 1 Triatominae as outgroup) showed positive and significant (p > 0.1) (Table 2). The level of significance on nucleotide diversity and the Tajima test statistic showed that neutrality analysis of nad1-nad2 genes (D = 0.925) were similar in significance compared to complete mitogenome (D = 0.931). There were a total of 14,975 (complete mitogenome) and 3,001 (nad1-nad2) positions in the final dataset with segregating sites

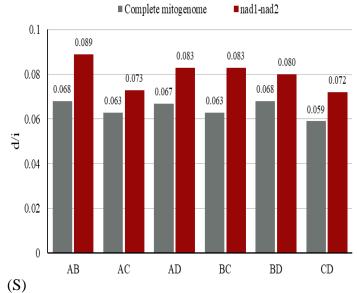


Figure 2. Tajima's relative rate test - Divergent sites(d)/Identical sites(i) where species A-D are Harpactorinae tested in combination with outgroup E - *Rhodnius prolixus**

6,049 and 1,303 for the complete mitogenome and nad1-nad2, respectively. High nucleotide diversity was observed for the (π) of 0.217 and 0.234 for complete mitogenome $(\pi = 0.217)$ and nad1-nad2 $(\pi = 0.234)$ with slight higher diversity in nad1-nad2. The high nucleotide diversity is a clue of distance genetic relationship of species studied in harpactorinae.

Table 2. Tajima's test of neutrality for the selected biocontrol agents mitogenome complete sequence and the sequence between *nad1-nad2* (*Rhodnius*

prolixus - Outgroup).

	Complete mitogenome	nad1-nad2
m	5	5
n	14975	3001
S	6049	1303
p_s	0.404	0.434
Θ	0.194	0.208
π	0.217	0.234
D	0.931*	0.925*

m= number of analysed sequences, n= total number of sites and S= Number of segregating sites. $p_s=S/n$, $\Theta=p_s/a_1$, $\pi=$ nucleotide diversity and D is the Tajima test statistic. * Significant (p > 0.1).

Relative rate test for evolution

Tajima's relative rate test was computed to test the equality of evolutionary rate between sequences AB, AC, AD, BC, BD and CD with outgroup sequence E (2 harpactorinae taxa with 1 outgroup triatominae). Results of the computation presented in the Figure 2 shows the Divergent sites(d)/Identical sites(i) form the relative rate test varies from 0.059 (CD) to 0.068 (AB and BD) for the complete mitogenome and 0.072 (CD) to 0.089

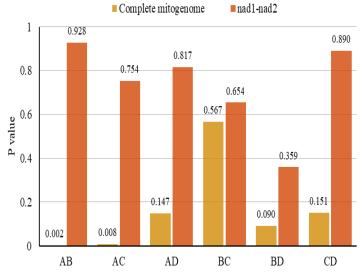


Figure 3. Tajima's relative rate test - P value, where species A-D are Harpactorinae tested in combination with outgroup E - *Rhodnius prolixus**. P<0.05 rejects null hypothesis for equality of evolutionary rate (AB) for the *nad1-nad2*.

The observed p value < 0.05 rejects null hypothesis for equality of evolutionary rate (Figure 3) at AB (0.002) and AC (0.008) for the complete mitogenome (Figure 3).

Phylogenetic tree

Similar trees were obtained using *nad1-nad2* (Figure 4 - left) and complete mitogenome (Figure 4 - right). Length of the branch from the outgroup is identical (0.16). Bootstrap estimates are low at 63 and 31 at the *nad1-nad2* corresponding to the high estimates at 100. *Agriosphodrus dohrni* is closely related to *Velinus nodipes*. However, the genetic distance between *Agriosphodrus dohrni* and outgroup Rhodnius prolixus was relatively far. *Rhynocoris fuscipes* is more similar to the

133

outgroup *Rhodnius prolixus*. Outline of phylogenetic trees using *nad1-nad2* region and mitogenome of *Agriosphodrus dohrni*, *Velinus nodipes*, *Scipinia*

horrida, Rhynocoris fuscipes and outgroup Rhodnius prolixus revealed that the tested predators showed high genetic distance and less genetic similarity among the tested harpactorinae (Figure 4).

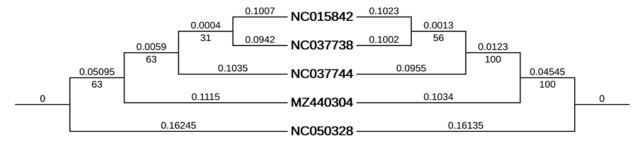


Figure 4. Phylogenetic tree constructed using nucleotide sequence between *nad1-nad2* (left) and complete mitogenome (right) with neighbourhood-joining method, Jukes-Cantor substitution model (200 bootstraps replications). NC015842: *Agrisphodrus dohrni;* NC037738: *Velinus nodipes;* NC037744: *Scipinia horrida;* MZ440304: *Rhynocoris fuscipes* and outgroup NC050328: *Rhodnius prolixus.*

DISCUSSION

The NCBI GenBank nucleotide database (https://www.ncbi.nlm.nih.gov/nuccore) provides <80 Reduviidae complete mitochondrial genomes and only 48 are archived as reference genomes (https://www.ncbi.nlm.nih.gov/refseq).

Considering the biocontrol potential, the complete mitogenomes of four harpactorinae predators viz., Agriosphodrus dohrni (Yao et al., Rhynocoris fuscipes, Scipinia horrida (Ambrose, 1999 and 2007) and Velinus nodipes (Takeno, 1998) were selected for the study. Selvaraj et al. (2022) reported the variable nucleotide sequences in complete mitogenomes of Triatominae and found the sequences between the genes cytB-nad1 and nad1-nad2 to be highly variable across ten species. This study is the next step in validating the pattern within harpactorinae biocontrol agents. The sequence length between the nad1-nad2 especially is highly variable ranging from 3,324 to 4,039 bp directly proportionate to the length of the complete mitogenome. The nucleotide sequence between cytB-nad1 encodes (5'-3') a non-coding RNA tRNA-Serine along with a few base pairs. Whereas the nucleotide sequence between nad1nad2 encodes complement (3'-5') tRNA-Leucine, 16s rRNA, tRNA-Valine, 12s rRNA and from 5'-3', a control region (miscellaneous feature), tRNA-Isoleucine, complement (3'-5') tRNA-

outgroup NC050328: Rhodnius prolixus. Glutamine and 5'-3' tRNA-Methionine (Cameron, 2014i & ii; Zhao et al., 2015; Chen et al., 2018; Zhao et al., 2020; Sun et al., 2021; Yi et al., 2022). The base composition is heavily based towards A and T which is common in the mitochondrial genomes of hemipterans, similar richness on A and T was observed in the tested mitogenome (Wang et al., 2015).

Phylogenetic analysis using the Tajima's test of neutrality revealed similarity in the statistic values. This shows that the changes occurring in the complete mitogenome is reflected at the nucleotide sequence between the genes nad1-nad2. In the relative rate test, the Divergent sites(d)/Identical sites(i) where species A-D are Harpactorinae tested in combination with outgroup E - Rhodnius prolixus shows that rate of evolution at nad1-nad2 is higher than the complete mitogenome i.e, it can be used as a molecular marker to validate the interspecific variations. Further, the P values observed in the complete mitogenome of AB and AC with outgroup E are significant (<0.05) showing equality in the evolutionary rate in contradiction to the values at nad1-nad2 where the P values are >0.05. Due to this phenomenon the nucleotide sequence between genes nad1-nad2 are unique to individual species (Nei and Kumar, 2000; Selvaraj *et al.*, 2022)

The phylogenetic tree constructed from the whole mitogenome coverage is similar to the one constructed using nad1-nad2. Therefore, the nucleotide sequence between nad1-nad2 can be considered to be a species-specific marker. Consequently, morphological systematics compulsory to validate the species (Trautwien, 2012) and the phylogenetic trees are an observable estimate of a long period of genome evolution which is owed to change with the calculation algorithm (Bickel, 2022). However, the molecular assembly can be influenced by the reference genome used to assemble the sequenced contigs. Hence, it is essential to validate the reports of this study across the subfamilies, genera and species for adoption of this variable sequence between nad1-nad2 for phylogenetic systematics.

In conclusion, the positive and significant nucleotide diversity on *nad1-nad2* genes and complete mitogenome revealed the high distance genetic and less genetic similarity among the species studied in Harpactorinae, *Agriosphodrus dohrni, Velinus nodipes, Scipinia horrida,* and *Rhynocoris fuscipes,* which clearly suggest the possible utilization of *nad1-nad2* genes as molecular marker to distinguish the species of Harpactorinae.

ACKNOWLEDGEMENTS

The authors appreciate Dr. Dunston P. Ambrose, CSIR Professor Emeritus and Founder, Entomology Research Unit (ERU) for his guidance with the taxonomy of Reduviidae.

REFERENCES

- Ambrose, D. P. 1999, Assassin bugs, Oxford and IBH publishing Co.Pvt.Ltd, Kolkata.
- Ambrose, D. P. 2007, The Insects: Beneficial and Harmful Aspects. Kalyani Publishers, New Delhi. p. 801.
- Ambrose, D. P. and Ganesh Kumar, A. 2016,
 Reduviid Predators, Ecofriendly Pest
 Management for Food Security, First Edition,
 PP. 217-257
- Barrion, A. T., Anguda, R. M., Litsinger, J. A., 1987, The natural enemies and chemical control of the Leucaena psyllid *Heteropsyila cubana*

- Crawford in the Phillipines. *Leucaena Research Report*, **7**:45–49
- Bickel, D. R. 2022, Phylogenetic Trees and Molecular Evolution: A Hands-on Introduction with Uncertainty Quantification Corrected. Springer Nature
- Cameron, S. L. 2014i, How to sequence and annotate insect mitochondrial genomes for systematics and comparative genomics research, *Systematic Entomology*, **39**:400-411.
- Cameron, S. L. 2014ii, Insect mitochondrial genomics: Implications for evolution and phylogeny, *Annual Review of Entomology*, **59**:95-117.
- Chen, J. Y., Chang, Y. W., Zheng, S. Z., Lu, M. X. and Du, Y. Z. 2018, Comparative analysis of the Liriomyza chinensis mitochondrial genome with other Agromyzids reveals conserved genome features. *Scientific reports*, **8**(1):1-10
- Du, Z., Liu, H., Li, H., Ishikawa, T., Su, Z. H., Cai, W., Kamitani, S. and Tadauchi, O., 2018. Invasion of the assassin bug *Agriosphodrus dohrni* (Hemiptera: Reduviidae) to Japan: Source estimation inferred from mitochondrial and nuclear gene sequences. *International journal of biological macromolecules*, 118:1565-1573.
- Jung, S., Duwal, R. K. and Lee, S. 2010, COI barcoding of True bugs, DNA Barcoding, *Molecular Ecology Resources*, **11**:266-270.
- Kjer, K., Borowiec, M. L., Frandsen, P. B., Ware, J. and Weigmann, B. M. 2016, Advances using molecular data in insect systematics, *Current Opinion in Insect Science*, **18**:40-47.
- Nei, M. and Kumar, S. 2000, Molecular Evolution and Phylogenetics. *Oxford University Press*, New York.
- Quicke, D. L. J. 1993, 'Principles and techniques of contemporary taxonomy', *Springer Science* + *Business Media*, B.V.
- Sahayaraj, K 2007, 'Pest control mechanism of Reduviids' (ISBN. 81-89473-03-04). Oxford Book Publication, Jaipur, **PP**. 240
- Sahayaraj, K 2014, 'Reduviids and their merits in biological control', Basic and applied aspects of

- biopesticides. New Delhi: *Springer India*, **10**:195-209
- Sahayaraj, K., Selvaraj, P. and Nirmala, K., 2002. Biological control potential of a reduviid predator, *Rhynocoris fuscipes* (Fab.) on three groundnut pests. *Asian J Microbiol Biotechnol Environ Sci*, **4**:451-455.
- Schuh, R. T. and Weirauch, C. 2020, True bugs of the world (Hemiptera: Heteroptera) Classification and natural history, *2nd edition*, *Siri Scientific Press*, UK.
- Selvaraj, P., Sherlin John, J., AbdulAzeez, S., Francis Borgio, J. and Ambrose, D. P. 2022, 'Variable nucleotide sequences between mitochondrial encoded protein-coding genes of *Triatominae* (Heteroptera: Reduviidae)', *Annual International Conference on Multidisciplinary Research and Innovation*, 8: 209-217
- Sun, Y., Huang, H., Liu, Y., Liu, S., Xia, J., Zhang, K. and Geng, J. 2021, 'Organization and phylogenetic relationships of the mitochondrial genomes of *Speiredonia retorta* and other lepidopteran insects'. *Scientific Reports*, 11(1):1-12
- Takeno, K., 1998. Enumeration of the Heteroptera in Mt. Hikosan, western Japan with their hosts and preys. *Esakia*. **38**:29–53
- Tamura K., Stecher G., and Kumar S. 2021. MEGA 11: Molecular Evolutionary Genetics Analysis Version 11. Molecular Biology and Evolution. https://doi.org/10.1093/molbev/msab120.
- Trautwien, M. D., Weigmann, B. M., Beutal, R., Kjer, K. M. and Yeates, D. K. 2012. Advances in Insect Phylogeny at the Dawn of Postgenomic Era. *Annual Review of Entomology*, **57**:449-468.
- Wang, Y., Chen, J., Jiang, L. Y. and Qiao, G. X. 2015. Hemipteran mitochondrial genomes: features, structures and implications for phylogeny. *International Journal of Molecular Sciences*, **16:**12382-12404.
- Weirauch, C., Bérenger, J. M., Berniker, L., Forero, D., Forthman, M., Frankenberg, S., Freedman, A., Gordon, E., Hoey-Chamberlain, R., Hwang, W. S., Marshall, S. A., Michael, A., Paiero, S.

- M., Udah, O., Watson, C., Yeo, M., Zhang, G. and Zhang, J. 2014. An Illustrated Identification Keys to the Reduviidae Subfamilies and Tribes, *Canadian Journal of Arthropod Identification*, No. 26.
- Yao, D., Yan, J., Li, G and Liu, H., 1995. A study on the morphology and bionomic of predatory bug *Agriosphodrus dohrni* (Signoret). *Forest Research.* 8:442–446.
- Yi, J., Wu, H., Liu, J., Li, J., Lu, Y., Zhang, Y., Cheng, Y., Guo, Y., Li, D. and An, Y. 2022. Novel gene rearrangement in the mitochondrial genome of *Anastatus fulloi* and phylogenetic implications for Chalcidoidea. *Scientific Reports*, 12(1):1-12
- Zhao, G., Li, H., Zhao, P. and Cai, W. 2015. Comparative mitogenomics of the assassin bug genus *Peirates* reveal conserved mitochondrial genome organization of *P. atromaculatus*, *P. fulvescens* and *P. turpis*, PLoS ONE, **10**(2), e0117862.
- Zhao, M. Y., Huo, Q. B. and Du, Y. Z. 2020. Molecular phylogeny inferred from the mitochondrial genomes of *Plecoptera* with *Oyamia nigribasis* (Plecoptera: Perlidae). *Scientific Reports*, **10**(1):1-10

Sherlin John, $J_{*}^{1,2}$, Selvaraj, P^{2*} ., Pushpanathan, T^{2} ., Ravichandran, B^{3} ., Sayed AbdulAzeez⁴ and Francis Borgio, J^{4} .

¹Research Scholar (18211282191036)

²Entomology Research Unit (ERU), Department of Zoology, St. Xavier's College (Autonomou), Palayamkottai-627002, Affiliated to Manonmaniam Sundaranar University, Tirunelveli-627012, Tamil Nadu, India.

³Department of Zoology, Sri S. Ramasamy Naidu Memorial College, Sattur, Virudhunagar-626203, Tamil Nadu, India

⁴Department of Genetic Research, Institute for Research and Medical Consultation (IRMC), Imam Abdulrahman Bin Faisal University, Dammam 31441, Saudi Arabia

*Corresponding author

E-mail: drselvabernad@gmail.com

Mobile: 0091 9791057506

Orcid ID: https://orcid.org/0000-0003-2257-7217