

# Antioxidant property of fresh and marine water cyanobacterial extracts in Swiss mice

R. Navanietha Krishnaraj<sup>1</sup>, S. Venkatesh Babu<sup>2</sup>, B. Ashokkumar<sup>3</sup>, P. Malliga<sup>4</sup> and P. Varalakshmi<sup>2\*</sup>

# ABSTRACT

Continuous usage of pesticide is the main cause of cellular damage by generation of free radicals. Antioxidants are intimately involved in the prevention of cellular damage. Hence the present investigation is mainly focused to study the antioxidant property of cyanobacterial extracts from diverse environments in order to prevent the free radicals toxicity. The alcoholic extracts of different cyanobacterial isolates including *Oscillatoria salina Synechcococcus, Oscillatoria annae, Oscillatoria chlorina, Spirulina sabsalsa and Spirullina platensis* were analyzed for their antioxidant property by physical body weight change, swimming time and biochemical parameters (superoxide dismutase activity and total reduced glutathione activity) by using Swiss mice. Stress was induced by forced swimming test and the antioxidant efficiency of cyanobacterial extracts was determined. The results showed that *Spirulina platensis* possess significant antioxidant property and *Synechcococcus sp* possess least activity when compared to other cyanobacterial isolates and control.

Key words: Body weight, cyanobacteria, reactive oxygen radicals, super-oxide dismutase

# INTRODUCTION

Pesticides are toxic; they are also potentially hazardous to humans, animals and other living beings in the environment. In this present scenario the continuous exposure to pesticides causes severe cellular and molecular damage to humans and other animals by generating free radicals. Antioxidants are substances or nutrients in our food which can prevent or slow the oxidative damage to our body. Phytonutrients and pigments present in the cyanobacteria act as antioxidants which facilitate the formation of the body's defense against free radical damage to cells. Antioxidants act as free radical scavengers and prevent and repair damage done by the free radicals. Reactive oxygen species (ROS) are often generated either as byproducts of biological reactions or from exogenous factors (Cerutti, 1991). It includes superoxide radicals, hydroxyl radicals, singlet oxygen, and hydrogen peroxide. ROS generally play a positive role such as energy production, phagocytosis, regulation of cell growth and intercellular signaling, or synthesis of biologically important compounds (Halliwell, 1997). But, ROS may also play a negative role; they can attack lipids in cell membranes and also attack DNA, inducing oxidations that cause membrane damage such as membrane lipid peroxidation and a decrease in membrane fluidity, and also cause DNA mutation leading to cancer (Pietta, 2000). An antioxidant is a substance that present at low concentrations compared to an oxidizable substrate has the ability to prevent or delay different types

© JBiopest. 320

of cell damage. The antioxidant defense mechanisms in biological systems are of two types namely enzymatic and non-enzymatic reactions. The enzymatic antioxidants include catalase and hydroperoxidase. The non enzymatic antioxidants include nutrient antioxidants like carotenoids, *a* tocopherol, ascorbic acid, glutathione, flavonoids, uric acid and plasma proteins such as transferrin, albumin, metalothionein etc. (Luximon Ramma *et al.*, 2002; Serena *et al.*, 2010).

There is a great demand throughout the world in finding new natural sources for antioxidants to prevent oxidative damage to living cells and to reduce the deterioration of food by oxidation (Pratt, 1992). Traditionally, some antioxidants such as tea, wine, fruits, vegetables and spices are used from the ancient days. Cyanobacteria are prokaryotic organism contains a wide variety of antioxidant pigments than the plants and most algal source (Robbins, 1987). Screening of cyanobacteria for antibiotics and other pharmacologically active compounds, has received ever-increasing interest as a potential source for new drugs. Cyanobacteria are known to produce metabolites with diverse biological activities such as antibacterial (Jaki et al., 2000), antifungal (Kajiyama et al., 1998), antiviral (Patterson et al., 1994), anticancer and antiplasmodial activity (Papendorf et al., 1998). Recently antioxidant property of cyanobacteria especially from O.annae has been reported by Rajavel et al. (2011). Carotenoids are the most widely distributed and structurally diverse classes of natural pigments predominantly

#### Navanietha Krishnaraj et al.

produced by cyanobacteria and that are doing important functions in photosynthesis and nutrition. Also they have potent anti oxidant activity. With this background of this present study mainly focused to screen are the antioxidant property of five different cyanobacterial isolates like *Oscillatoria annae*, *O. chlorina*, *Spirullina sabsalsa*, *Synechococcus and S. platensis* in order to prevent the oxidative damage caused by the pesticides because that are absorbed on the surface of vegetables and fruits would cause severe damage to the health of the animals while they consume the fruits and vegetables.

#### MATERIALS AND METHODS

Swiss mice were the animal model used for this experiment. Mid log phase culture of different cyanobacterial isolates including O. salina, Svnechcococcus, O. annae, O. chlorina, S. sabsalsa and S. platensis were collected from National Facility for Marine Cyanobacteria, Bharathidasan University, Tiruchirappalli. The cultures were grown BG11 and ASN medium in a culture flask separately. The cultures were allowed to grow till they reached the mid log phase. Five different strains of cyanobacteria (1 g fresh weight) were homogenized separately with glass powder and 75% alcohol using Mortar and Pestle. The homogenized extracts were centrifuged at 5000 rpm for 10 minutes. The clear extract was separated and dried using speed vac concentrator. Antioxidant effect of different cyanobacterial extracts were analyzed by measuring the level of antioxidant activity before and after the stress induction to the experimental animal on 1st, 14th and on 28th day.

#### Swimming test

Stress was induced by forced swimming test. Induction of Stress (Nagaraja and Jeganathan, 1999) was carried out in polypropylene tub 90 cm height, 90 cm diameter and 60 cm depth of water. The water was maintained at 18°C by adding ice cubes to the container. Male albino rats of Swiss strain (130 to 200g) were isolated into 19 groups and each group contains 6 animals.

# Analysis of superoxide dismutase and total reduced glutathione activity

Animals were examined carefully, weighed and placed at room temperature (30°C) in normal environmental conditions. They were fed with normal diet (pellet) directly into the oesophagus using curved feeding tube daily at 11:00 am. On 1st, 14th and 28<sup>th</sup> days the animals were weighed and were given stress. The blood samples (2 mL) were taken for the analysis of antioxidant effect by puncturing the retro orbital plexus directly into heparinised micro capillary tube into a test tube containing 0.1 ml of heparin. The physiological parameters (bodyweight changes), the biochemical parameters super oxide dismutase activity in haemolysate (Marklund and Marklund, 1974) and total reduced Glutathione activity in haemolysate (Patterson and Lazarow, 1975; Gul and Kutay, 2000) were analyzed.

#### **Experimental animal groups**

Five groups of animals were used for this study. They were: Group A - A1 = 0.5 µg/L of *Spirullina sabsalsa*; A2 = 1.0 µg/L of *Spirullina sabsalsa*; A3 = 1.5 µg/L of *Spirullina sabsalsa*; Group B - B1 = 0.5 µg/L of *Synechococcus*; B2 = 1.0 µg/L of *Synechococcus*; B3 = 1.5 µg/L of *Synechococcus*; Group C-C1 = 0.5 µg/L of *Spirullina platensis*; C2 = 1.0 µg/L of *Spirullina platensis*; C3 = 1.5 µg/L of *Spirullina platensis*; Group D - D1 = 1.0 µg/L of *Oscillatoria annae*; D2 = 1.0 µg/L of *Oscillatoria annae*; D3 = 1.5 µg/L of *Oscillatoria annae*; Group E - E1 = 0.5 µg/L of *Oscillatoria chlorina*; E2 = 1.0 µg/L of *Oscillatoria chlorina*; E3 = 1.5 µg/L of *Oscillatoria chlorina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillatoria salina*; F2 = 1.0 µg/L of *Oscillatoria salina*; F3 = 1.5 µg/L of *Oscillator* 

#### **RESULTS AND DISCUSSION**

# Total reduced glutathione activity

The biological antioxidant system has several enzymes to protect the body from free radicals. Reduced glutathione is one of the enzymes which can be considered as a marker for antioxidant activity. Reduced glutathione is directly proportional to the amount of biological antioxidant activity. GSH based antioxidant study shows that *S. platensis* has higher reduced glutathione activity when compared with *O. salina*, *O. annae*, *O. chlorina Synechococcus* sp and *S. sabsalsa*. *O. chlorina* and *Synechococcus* sp has the least GSH activity. *O. salina*, *O. annae* and *S. sabsalsa* have intermidiate activity. So the experiment reveals that *S. platensis* extract may prevent the oxidative damage caused by pesticides.

#### **Body weight changes**

A body weight change is one of the physical parameters to study the oxidative stress. Induction of stress increases the body weight of mice. On supplementing the antioxidants, the body weight decreases. The body weight has significantly reduced in *S. platensis* when compared to *O. salina*, *O. annae*, *O.chlorina* and *S. sabsalsa*. Obviously, *S. platensis* has higher antioxidant activity. *O. chlorina* and *Synechococcus* sp. has least GSH activity. *O. salina*, *O. annae* and *S. sabsalsa* have intermidiete activity.

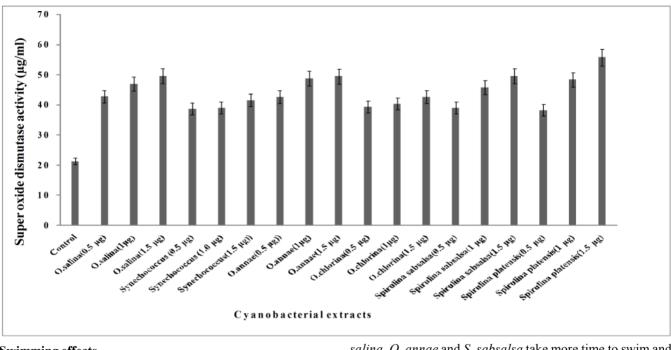


Figure 1. Effect of cyanobacterial extracts in the total reduced glutathione of Swiss Albino mice

# Swimming effects

Swimming time is one of the physical parameters to study antioxidant activity. Induction of stress increases the swimming time of mice. Use of antioxidant decreases the swimming time. O. salina ( $0.5\mu$ g) has the lowest swimming time and has the highest antioxidant activity. S. platensis ( $1.5\mu$ g) also has a moderately higher antioxidant activity. O. *salina*, *O. annae* and *S. sabsalsa* take more time to swim and have least antioxidant activity.

# Analysis of Super Oxide Dismutase

Like reduced glutathione, SOD is another important natural free radical scavenging antioxidant enzyme. So, the amount of SOD expressed is directly proportional to the antioxidant activity. *S. platensis* has higher SOD activity. *Synechococcus* 

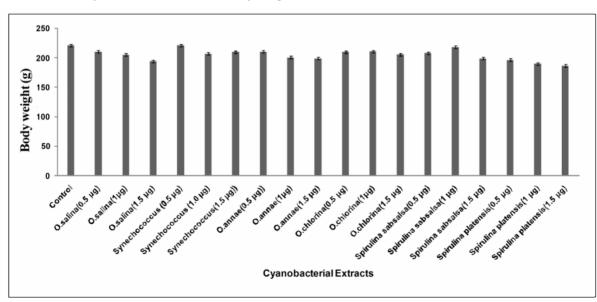
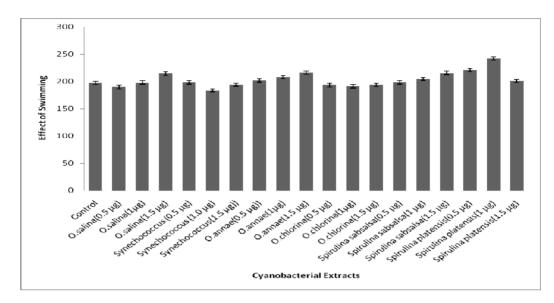
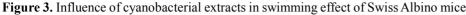


Figure 2. Influence of cyanobacterial extracts in body weight of Swiss Albino mice

252



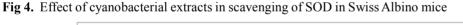


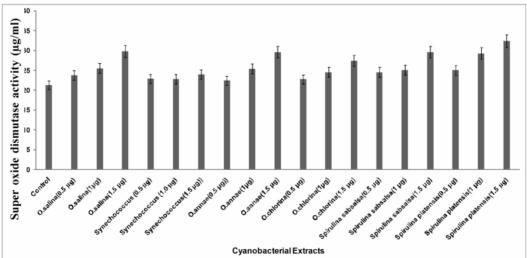
shows least SOD activity. *O. salina*, *O. annae*, *O. chlorina* and *S. sabsalsa* exhibit moderate SOD activity.

The extract of *S. platensis* has the potent anti oxidant activity in swiss albino mice. Hence *S. platensis* can be a new pharmaceutically valuable source for the animals ingested the with toxic pesticides in order to reduce the free radicals formation.

# REFERENCES

- Cerutti, P. A. 1991. Oxidant stress and carcinogenesis. European Journal of Clinical Investigation, **21:** 1-11.
- Gul, M. and Kutay, F. Z. 2000. Cellular and clinical implications of glutathione. *Indian Journal of Experimental Biology*, 38: 625-634.
- Halliwell, B. 1997. Antioxidants and human diseases: a general introduction. *Nutrition Review*, **55**: 44-52.
- Jaki, B., Heilmann J. and Sticher, O. 2000. New antibacterial metabolites from the cyanobacterium Nostoc commune (EAWAG 122b). Journal of Natural Products, 63: 1283-1288.
- Kajiyama, S., Kanzaki, H., Kawazu, K. and Kobayashi, A. 1998. Nostifungicidine, an antifungal lipopeptide from the field





#### Antioxidant property of cyanobacterial extracts

grown terrestrial blue-green alga, *Nostoc commune*. *Tetrahedron Letters*, **39**: 3737-3740.

- Luximon Ramma, A., Bahorun, T., Soobrattee, M. and Aruoma, O.I. 2002. Antioxidant activities of phenolic, proanthocyanidin and flavonoid components in extracts of *Cassia fistula*. *Journal of Agriculture Food Chemistry*, 50: 5042–5047.
- Marklund, S. and Marklund, G. 1974. Involvement of superoxide anion radical in autooxidation of pyrogallol and convenient assay for superoxide dismutase. *Europian Journal of Biochemistry*, **47**: 469-474.
- Nagaraja, H. S. and Jeganathan, P. S. 1999. Comparative study of different types of stress on some physiological and biochemical parameters in albino rats. *Biomedicine*, **19**(2): 137-149.
- Papendorf, O., König, G. M., Wright, A. D. and Hirridin, B. 1998. 2,4-dimethoxy-6 eptadecylphenol, secondary metabolites from the cyanobacterium *Phormidium ectocarpi* with antiplasmodial activity. *Phytochemistry*, **49**: 2383-2386.
- Patterson, J. W. and Lazarow, A. 1975. Methods of biochemical analysis. Interscience publishers inc. New York, 259 PP.
- Patterson, G. M. L., Larsen, L. K. and Moore, R. E. 1994. Bioactive natural products from blue-green algae. *Journal* of Applied Phycology, 6: 151-157.
- Pietta, P. G. 2000. Flavonoids as antioxidant. *Journal of Natural Products*, **63**: 1035-1042.
- Pratt, D. E. 1992. Natural antioxidants from plant material, Phenolic compounds. In: Food and their effects on health. American Chemical Society, Washington, (ACS Symposium Series, 507): 54-71.

- Rajavel, R., Sivakumar, T., Jagadeeswaran, M., Rajesh, V. and Malliga, P. 2011. Evaluation of *in vitro* and *in vivo* antioxidant activity of Oscillatoria annae. The Internet Journal of Pharmacology, 9 (2).
- Robbins, J. 1987. Anti-inflammatory and Antioxidant effects. Diet for A new America: 20.
- Schuler, P. 1990. Natural antioxidants exploited commercially, In: *Food Antioxidants* (Hudson, B. J. F. ed.). Elsevier, London. 99 -170 PP.
- Serena, M. M., Balasubramani, M., Rajan, K. and Gerald, I. A. J. 2010. Evaluation of the larvicidal activity of the leaf extracts of *Duranta erecta* Linn. (Verbenaceae) on the larvae of *Culex quinque fascitatus* (Say) (Culicidae). *Journal of Biopesticides*, 3(3): 582-585.

**R. Navanietha Krishnaraj<sup>1</sup>, S. Venkatesh Babu<sup>2</sup>, B. Ashokkumar<sup>3</sup>, P. Malliga<sup>4</sup> and P. Varalakshmi<sup>2\*</sup>** <sup>1</sup>Department of Biotechnology, Kalasalingam University, Krishnan Koil, Tamil Nadu, India.

<sup>2</sup>Department of Molecular Microbiology, School of Biotechnology, Madurai Kamaraj University, Madurai, Tamil Nadu, India.

<sup>3</sup>Department of Genetic Engineering, School of Biotechnology, Madurai Kamaraj University, Madurai, Tamil Nadu, India.

<sup>4</sup>Department of National Facility Marine Cyanobacteria, Bharathidasan University, Tiruchirappalli, Tamil Nadu India.

\*E.Mail: vara5277@gmail.com

Received: September 04, 2011

Revised: October 10, 2011

Accepted: February 12, 2012