

Bioefficacy of *Trichoderma* isolates against pathogenic fungi inciting spinach (*Spinacea oleracea* I.)

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ABSTRACT

Biological control of plant pathogens using antagonistic fungi can be successfully utilized especially within the frame work of integrated disease management system. Efficacy of different isolates of Trichoderma species against two pathogenic fungi causing leaf spot (Alternaria spinaciae) and wilt (Fusarium oxysporum f. sp. spinaciae) on spinach (Spinacea oleracea L.) by dual culture technique under in vitro conditions. Generally Trichoderma used as a biological control for the inhibition of growth of the pathogens. Different isolates i.e. Trichoderma viride (12), T. harzianum (12), T. koningii (07), T. pseudokoningii (05) and T. virens (10) strains were isolated and used for antagonistic study. Results indicated that, among T. viride isolates, Tv_1 and Tv_5 showed maximum antagonism, but in case of Tv_7 and Tv_{10} showed minimum antagonism against A. spinaciae. In case of F. oxysporum f. sp. spinaciae showed significant inhibition by Tv₄ and reduced the percent inhibition by Tv₈ and Tv₆. In case of *T. harzianum* isolates, Th₃ and Th₁₀ showed maximum inhibition on A. spinaciae while in F. oxysporum f. sp. spinaciae, Th₅ increased the inhibition and decreased in Th₃ and Th₇. Trichoderma koningii showed better results in Tk₇ isolate whereas decreased Tk₄ over A. spinaciae, but in F. oxysporum f. sp. spinaciae reduced rate was more in Tk₄ and Tk₂. Isolates of *T. pseudokonongii* found maximum inhibition in Tp₅ over *A. spinaciae* and significant inhibition in Tp₃ over F. oxysporum f. sp. spinaciae. Trichoderma virens isolates were found significant inhibition in Tvr₄ and Tvr₉ and 50% reduction over A. spinaciae and Tvr₄ increased the percent inhibition and decreased the Tvr₇ in case of F. oxysporum f. sp. spinaciae.

Key words: Alternaria spinaciae, dual culture, Fusarium oxysporum spinaciae, Trichoderma

INTRODUCTION

Spinach (Spinacea oleracea L.) is an important vegetable crop. Although many diseases have been reported as externally and internally seed borne (Correl and Monelok, 1994; Sumner, 1991; Walker, 1952; Bhale, 2002; Raid and Kucharek, 2003) which causes discolouration, spoilage of seeds, germination effects, rotting and wilting of seedlings, stem gall, blight, leaf spot, powdery mildew, downy mildew, rust and anthracnose. Out of which leaf spot (Alternaria spinaciae Allesch and Noack) and wilt (Fusarium oxysporum f. sp. Spinaciae (Sherb) Synder and Hansen) diseases to be severe. Since the diseases are soil and seed borne, is difficult to manage. Only practical solution seems to be use of resistant variety. However, resistant varieties for these diseases are not available. In this situation crop loss due to these diseases is substantial as it leads to capital losses up to 20 to 60%. Since, biocontrol agents

for protection of seeds and control of seed borne diseases offers farmers an alternative source of chemical fungicides which is highly effective (Callan et al., 1997). Fungicides such as metalaxyl, are however expensive and growers tend to use lower than optimum dosages (Csoinos and Bertrand, 1994). Furthermore, the use of fungicides, besides being expensive and involving risks to the environment associated with the application of chemicals, is not totally effective and may lead to the appearance of new, resistant strains of pathogens (Bruin and Edgington, 1980). *Trichoderma* has been the focal point of a number of studies concerning their ability to control plant pathogens. Strains of the species T. harzianum has shown effectiveness when used in disease control caused by several fungi, including Sclerotium rolfsii, a widely distributed and highly destructive plant pathogen (Benhamou and Chet, 1996), and Sclerotinia sclerotiorum affecting runner beans (Inbar et al. 1996). In a similar fashion, strains of

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T. harzianum, *T. koningii*, and *T. longibrachiatum* have been effective in *Armillaria* control in tea (Osando and Waudo, 1994), while *T. koningii* has been used for *Fusarium solani* corn rot control (McAllister *et al.* 1994).

It is therefore necessary to develop alternative ways of control. One such alternative is biological control, in which microorganisms are selected for their ability to antagonize pathogens. Therefore an investigation was made to evaluate the different isolates of *Trichoderma* species against the pathogenic fungi on spinach.

MATERIAL AND METHODS

Isolation of *Trichoderma* spp

Rhizosphere soils of irrigated and non irrigated plants were collected from different parts of Marathwada region of Maharashtra state. From the rhizosphere soil samples, desired strain of Trichoderma species were isolated by using potato dextrose agar (PDA) and Trichoderma selective medium (TSM) by dilution plate technique. The isolated strains were identified by reculturing on another petriplates containing sterilized TSM. The isolated strains were identified up to species level based on colony characters, growth of fungus, structure of conidiophores mycelium, and conidia. All Trichoderma spp. fungi were purified by hyphal tip technique and identified on the basis of cultural, microscopic and morphological characters (Kubicek and Harman, 2002). The isolated strains of Trichoderma species were maintained on PDA and TSM slants. This was maintained throughout the study by periodical transfers on (PDA) medium under aseptic conditions, to keep the culture fresh and viable.

Isolation of test pathogens

Test fungi *A. spinaciae* and *F. oxysporum* f. sp. *spinaciae* were isolated from the diseased part viz leaf spot and wilt of spinach (*Spinacea oleracea* L.) respectively Earlier, the pathogencity of test fungi were confirmed in inoculating spinach seedlings properly. The isolated test fungi were purified on aseptic conditions and maintained on PDA slants.

Dual culture experiment

Antagonistic efficacy of different isolates of Trichoderma species, T. viride, T. harzianum, T. koningii, T. pseudokoningii and T. virens were tested against the isolated pathogenic test fungi, dual culture experiment was described by Morton and Stroube, (1955). Trichoderma species and test fungi were inoculated at 6 cm apart. Three replications were maintained for each treatment and incubated at $27 \pm 2^{\circ}$ C for 7 days. Monoculture plates of both are served as control. After 7 days, radial growth of test fungi and Trichoderma isolates were measured. Colony diameter of test fungi in dual culture plate was observed and compared with control. Antagonistic effect of *Trichoderma* spp., as decrease of the mvcelial growth of pathogenic fungi.was determined using the Vincent (1947) method.

RESULTS AND DISCUSSION

Isolation of *Trichoderma* **species test pathogens** Five species of *Trichoderma*, T. *viride(12), T. harzianum(12), T. koningii(07), T.pseudokoningii* (05) and *T. virens(10)* were isolated from irrigated and non irrigated rhizosphere soil of Marathwada region. Perusal of results indicated that *A. spinaciae* and *F. oxysporum* f. sp. *spinaciae* were significantly influenced by various isolates of *Trichoderma* species under study at 7 days after inoculation (DAI).

Dual culture experiment

Table 1 indicated that *T. viride* isolates on growth of *A. spinaciae* Tv_1 and Tv_5 showed maximum antagonism as compared to others but Tv_7 and Tv_{10} decreased the antagonism. While in case of *F. oxysporum* f. sp. *spinaciae* showed greater inhibition by Tv_4 followed by Tv_3 , but Tv_8 and Tv_6 reduced the percent inhibition. *Trichoderma harzianum* isolates on growth of *A. spinaciae* were found more percent inhibition in Th₃ and Th_{10} . In case of *F. oxysporum* f. sp. *spinaciae* increased the percent inhibition in Th₅ whereas decreased in Th₃ and Th₇ (Table 2).

Trichoderma koningii isolates on growth of *A. spinaciae* showed more antagonism in Tk_7 but decreased in Tk_4 . But in case of *F. oxysporum* f. sp. *spinaciae* found more percent inhibition in Tk_3 (67.50%) as compared to other but reduced the percent inhibition Tk_4 and Tk_2 (Table 3).

Table 1. Influence of *Trichoderma viride* isolates on growth of *Alternaria spinaciae* and *Fusarium* oxysporum f. sp. spinaciae

Isolates	Locations	Radial growth of A.spinaciae (mm) (7DAI)	Inhibition %	Radial growth of F.oxysporum f. sp.spinaciae. (mm) (7DAI)	Inhibition %
Tv_1	Naldurg	14.00	84.44	40.22	55.50
Tv_2	Osmanabad	17.20	81.58 ^I	27.30	69.66
Tv_3	Latur	22.30	75.22	12.50	86.11
Tv_4	Nanded	35.10	61.00	09.00	90.00 ^I
Tv_5	Jalna	16.95	81.16 ¹	44.23	50.85
Tv_6	Aurangabad	27.00	70.00	55.15	38.72 ^D
Tv_7	Beed	55.15	38.72 ^D	27.00	70.00
Tv_8	Paranda	25.10	72.11	61.00	32.22 ^D
Tv_9	Ashti	18.95	78.94	47.60	47.12
Tv_{10}	Omerga	50.30	44.12 ^D	29.00	67.77
Tv_{11}	Ahmedpur	36.35	56.61	69.11	23.21 ^D
Tv_{12}	Hingoli	27.50	69.44	47.60	47.11
SE±			4.28		5.96
CD(P=0.05)			9.43		13.14

DAI- Days after inoculation, I - Increased percent inhibition, D- Decreased percent inhibition

Table 2. Influence of *Trichoderma harzianum* isolates on growth of *Alternaria spinaciae* and *Fusarium*oxysporum f. sp. spinaciae

Isolates	Locations	Radial growth of A.spinaciae (mm) (7DAI)	Inhibition %	Radial growth of F.oxysporum f. sp. spinaciae. (mm) (7DAI)	Inhibition %
Th_1	Tuljapur	20.12	77.70	30.12	66.10
Th_2	Kallam	34.12	62.08	37.10	58.77
Th_3	Ausa	45.00	50.00	57.00	36.67
Th_4	Nanded	12.00	86.66 ¹	61.37	31.81 ^D
Th_5	Badnapur	41.17	54.25	17.42	80.64 ^I
Th_6	Beed	22.39	75.12	35.00	61.11
Th ₇	Paithan	24.11	73.21	50.23	44.18
Th ₈	Paranda	27.67	69.25	29.14	67.62
Th ₉	Patoda	35.00	61.11	47.17	47.58
Th_{10}	Nilanga	16.00	82.22 ^I	24.27	73.03
Th_{11}	Udgir	34.00	62.22	29.00	67.77
Th_{12}	Parbhani	40.22	55.31	55.52	38.3 ^D
SE± CD(P=0.05)			3.38 7.44		4.59 10.11

DAI- Days after inoculation, I - Increased percent inhibition, D- Decreased percent inhibition

At 7 days after incubation (DAI), Т. pseudokoningii isolates on growth of A. spinaciae was observed maximum in Tp_5 and in case of F. oxysporum f. sp. spinaciae was found maximum in Tp₃ (Table 4). Table 5 illustrated that T. virens isolates on growth of A. spinaciae, maximum percent inhibition showed Tvr₄ and Tvr₉ but 50% inhibition was found in case of Tvr7. Maximum percent inhibition was observed in Tvr₄ and minimum in Tvr₇ in case of F. oxysporum f. sp. spinaciae.

Among all isolates of *T.viride*, isolate no 1 for *A*.

spinaciae and isolate no 4 for *F. oxysporum f. sp. spinaciae* was found suitable for antagonism. In *T. harzianum*, isolate no 4 for *A. spinaciae* and isolate no 5 for *F. oxysporum f. sp. spinaciae* was found maximum percent inhibition. In case of *T. koningii*, isolate no 7 for *A. spinaciae* and isolate no 3 for *F. oxysporum f. sp. spinaciae* was found significant antagonism. In *T.pseudokoningii* isolate no 5 for *A. spinaciae* and isolate no 3 for *F. oxysporum f. sp. spinaciae* was found significant antagonism. In *T.pseudokoningii* isolate no 5 for *A. spinaciae* and isolate no 3 for *F. oxysporum f. sp. spinaciae* was observed bioefficacy. In *T.virens* isolates no 4 and 9 were suitable for antagonism but maximum percent inhibition was failed in *F. oxysporum f. sp. spinaciae*.

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Isolates	Locations	Radial growth of A.spinaciae (mm) (7DAI)	Inhibition %	Radial growth of F.oxysporum f. sp.spinaciae. (mm) (7DAI)	Inhibition %
Tk_1	Naldurg	26.00	71.10	35.00	61.10
Tk_2	Osmanabad	27.12	69.86	50.11	44.32 ^D
Tk ₃	Latur	37.00	58.88	29.25	67.50
Tk_4	Aurangabad	51.50	42.77 ^D	57.44	36.17 ^D
Tk ₅	Jalna	28.11	68.76	39.39	56.23
Tk_6	Parbhani	32.35	64.05	38.57	57.14
Tk ₇	Nanded	12.33	86.30 ¹	31.11	65.43
SE -			5.01		11.87
SE± CD(P=0.05)			4.29		10.18

Table 3. Influence of *Trichoderma koningii* isolates on growth of *Alternaria spinaciae* and *Fusarium* oxysporum f. sp. spinaciae

DAI- Days after inoculation, I - Increased percent inhibition, D- Decreased percent inhibition

Table 4. Influence of *Trichoderma pseudokoningii* isolates on growth of *Alternaria spinaciae* and *Fusariumoxysporum* f. sp. *spinaciae*

Isolates	Locations	Radial growth of A.spinaciae (mm) (7DAI)	Inhibition %	Radial growth of F.oxysporum f. sp.spinaciae. (mm) (7DAI)	Inhibition %
Tp ₁	Ausa	23.00	74.40	35.00	61.11
Tp ₂	Tuljapur	29.25	67.50	37.25	58.61
Tp ₃	Jalna	22.11	75.43	24.00	73.33
Tp_4	Sillod	39.15	56.50	47.00	47.77 ^D
Tp ₅	Ardhapur	17.00	81.11^{I}	35.00	61.11
SE±			1.67		1.67
CD(P=0.05)			4.65		4.65

DAI- Days after inoculation, I - Increased percent inhibition, D- Decreased percent inhibition

Table 5. Influence of *Trichoderma virens* isolates on growth of *Alternaria spinaciae* and *Fusarium* oxysporum f. sp. spinaciae

Isolates	Locations	Radial growth of A.spinaciae (mm) (7DAI)	Inhibition %	Radial growth of F.oxysporum f. sp.spinaciae. (mm) (7DAI)	Inhibition %
Tvr ₁	Naldurg	24.00	74.00	27.00	70.00
Tvr ₂	Murum	24.15	73.16	30.11	66.54
Tvr ₃	Beed	29.11	67.65	34.39	61.78
Tvr_4	Badnapur	12.00	86.66 ¹	19.22	78.64
Tvr ₅	Kannad	32.00	64.44	41.11	54.32
Tvr ₆	Parbhani	21.37	76.25	33.13	63.18
Tvr ₇	Nilanga	47.11	47.65 ^D	50.11	44.32 ^D
Tvr ₈	Udgir	27.35	69.61	34.11	62.10
Tvr ₉	Nanded	12.00	86.66 ¹	33.12	63.20
Tvr_{10}	Hingoli	31.34	65.17	28.05	68.83
SE±			3.60		2.91
CD(P=0.05)			8.13		6.59

DAI- Days after inoculation, I - Increased percent inhibition, D- Decreased percent inhibition

Several workers has been reported that the use of *Trichoderma* species against number of plant pathogenic fungi. *Trichoderma viride* and *T. koningii* were found effective against *Rhizoctonia solani* (Mukhopadhyay, 1987), *T. hamatum* against *R. solani* (Lewis *et al.*, 1990). In *in*

vitro, control of *R. solani, F. oxysporum* and *Macrophomina phaseolina* were achieved with *T. koningii, T. hamatum* and *T. harzianum* (Arora, 1990). *Trichoderma harzianum* and *T. viride* were antagonistic to *F. oxysporum* f. sp. *lycopersici* (Gaikwad *et al.*, 1999) and to *R. solani* (Yadav and Tripathi, 1999). Recently, Waghmare and

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Kurundkar (2011) reported efficacy of *Trichoderma* species against *F. oxysporum* f. sp. *carthami* causing wilt of safflower and found that isolates no 29 *and*33 were found minimum growth of the pathogen as compared to others. The species of *Trichoderma* significantly inhibited the mycelial growth of plant pathogens (Rajkonda *et al.*, 2011).

According to Papavizas and Lumsden (1980) the mechanisms involved in the control of pathogens by Trichoderma spp. are probably: antibiosis, lysis, competition and mycoparasitism. However, Avers and Adams (1981) indicated that interactions observed in vitro do not necessarily confirm their operation for the decrease in pathogen populations and reduction in diseases observed in natural conditions. Further investigations are needed in order to characterize the antagonist-host interactions observed during these studies.

The *in vitro* screening with our arbitrary system of bio-antagonists effective against soil borne pathogens is a simplistic approach to understand a small sector of biological system in diseases control. Therefore, it may be more prudent to search for biological antagonists against specific pathogen and evaluate blends of antagonists for wider applications (Baker and Cook, 1974). Our results show that although considerable success in biocontrol is achieved under laboratory conditions the outcome is not proportionate under field conditions. Hence, work is needed towards a understanding and development better of technologies that allow the biocontrol agent to spread and proliferate in soil. In addition as suggested by Papavizas (1985) research should be directed towards the improvement of strains of biological agents that are more capable of becoming established and surviving under adverse field conditions. Thus, it is obvious that biological control offers, durable environmentally safe and cost effective alternative to chemical for the efficient management of plant disease

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