Field evaluation of *Pseudomonas fluorescens* against the pink bollworm, *Pectinophora gossypiella* and the spiny bollworm, *Earias vitella*.

Manjula^{1*} T. R., Kannan, G. S.² and Sivasubramanian, P.³

ABSTRACT

Pseudomonas fluorescens and *Beauveria basianna* were evaluated in cotton growing season against the common bollworms that infested cotton, pink bollworm, *Pectinophora gossypiella* and the spiny bollworm, *Earias vitella*. Reduction in the infestation and larval content was determined after three successive sprays with 15 days interval. The indirect effect of *Pseudomonas fluorescens* on seed cotton yield was considered and compared with the untreated check. Results of the present study revealed that the foliar application of *P. fluorescens* and *Beauveria basianna treatment* exhibited the greatest reduction in bollworms infestation. Soil and Foliar application of *P. fluorescens*, was recorded with the highest amount of seed cotton yield.

Keywords: Pseudomonas fluorescens, Beauveriabasianna and Eariasvitella.

MS History: 21.03.2018 (Received)-15.05.2018 (Revised)-26.05.2018 (Accepted).

Citation: Manjula T. R., Kannan G .S. and Sivasubramanian P. 2018. Field evaluation of *Pseudomonas fluorescens* against the pink bollworm, *Pectinophora gossypiella* and the spiny bollworm, *Eariasvitella*. *Journal of Biopesticides*, **11**(1): 25-29.

INTRODUCTION

Cotton Gossypium spp. (Family: Malvaceae) is one of the most economic non-field crops of global importance and main cash crop in the world. In India it is cultivated in 105 lakh hectares with production of 351 lakh bales of seed cotton. Moreover, due to the top most position in Indian Agriculture it is also popularly known as white gold. Cotton fibre is an important raw material for textile industries and plays a key role in national economy in term of employment generation and foreign exchange. Production of cotton is limited by various factors among which insect pests are very important. During growth period 148 insect pests have been recorded on cotton crop out of which only 17 species have been recorded as major insect pests of cotton crop. Cotton pests can primarily be divided into sucking pests and bollworm. Among the bollworm the pink bollworm, Pectinophora goeeypiella (Saund) and spiny bollworm, Earias vitella (Boisd.) are the most serious cotton pests (Hussein et al., 2001). Pests are such a serious threat to cotton production that

the cost of cotton pest control is about \$12.5 million (Younis et al., 2007). Bollworms have caused the greatest yield losses in nearly one million hectares cultivated annually in the world. (Haque, 1991; EI-Nagger, 1998). The reduction in cotton yield was mostly related to the late season infestation with both species and the economic yields are almost impossible to achieve without their chemical control. For both species most of larvae live inside the green bolls and pesticides used must be carefully selected to affect egg and adult stages with minimum side effect on beneficial arthropods. While searching for the best alternative use of bioinaculants and botanicals will be a good choice for controlling insect pests and they pave the way for eco-friendlier pest management. Bioinaculants of microbial controls an essential component in biointensive pest management, help reduce the dependence on chemical pesticides and ecological deterioration and serve as The utilization of plants owns insecticides. defense mechanism is the subject of current interest in the management of pest. Induced protection of plants against various pests and

T. R. Manjula et al.,

diseases by biotic and abiotic inducers has been reported in many crops. Plant growth promoting rhizobacteria (PGPR) has been shown to be capable of inducing pest resistance in addition to promoting plant growth. The main objective of the present study was to evaluate in - situ efficacy of *Pseudomonas fluorescens* against bollworm infestation and the reduction in the infestation must be associated with the greatest cotton yield

MATERIALS AND METHODS

Field experiments were conducted during 2014 -2015 at Vanavarayar Institute of Agriculture, Pollachi, Coimbatore District. P. fluorescens was evaluated against the two common bollworm infested cotton, *Pectinophora* gossypiella and Eariasvitella. The experiment was laid out in randomized block design (RBD). There were six treatments viz., T1- Foliar application of P. fluorescens @1%, T2 -Soil application of P. fluorescens 2.5 kg/ha, T3. - Soil and Foliar application of @1%. T4 P. fluorescens - Foliar application of P. fluorescens @1% and @ 1%, T5 - Foliar application of *B. basianna* @ 1%, T6 - Profenophos 50 EC @ 1 lit/ha. along with a T7 - control treatment. Each treatment was replicated four times. The plot size of each experimental unit was 6 x 5 m. Row to row and plant to plant distance was maintainex as 90 x 60 cm. The percent infestation was calculated by the following formula:

No. of green bolls damaged

% infestation: -----Total no. of bolls

Biweekly pests counting was carried out before and after the treatment spray upon attainment of economic threshold level (ETL) of both pink boll worm and spotted bollworm infestation (5 larvae/ 25 plants or 10% infestation of fruiting bodies). Treatments were sprayed according to their label recommended dose with the help of knapsack hand sprayer early in the morning using hollow- cone nozzle. Samples of 100 green bolls per treatment (25 bolls for each treatment) were taken at random and dissected. For each treatment, reduction percentages in bollworm infestation, bollworm larval content were calculated using Henderson and Titlon (1955) equation :

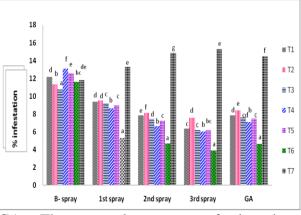
Per cent Reduction = [1- { (Control before* treatment after) / (Control after* treatment before) }]* 100.

The seed cotton yield for each plot was harvested and weighed then mean weight of seed cotton yield was compared among the treatment and the untreated check. Data were analyzed using analysis of variance followed by Tukey's multiple comparison tests (Gonez and Gonez, 1984).

RESULTS

The pre spray infestation of both species ranged between 10-13%. Six weeks after application, the infestation averaged 3.86 to 7.54 % in *P. fluorescen s*and *B. basianna* treatments compared to 15.25 % in control treatment (fig.1).

Fig. 1. Percentages of bollworms infestation before and after applying *P. fluorescens* treatments in three successive sprays with 15 days interval. ANOVA, Tukey's at 0.05 level of probability; same alphabets during a counting indicates insignificant

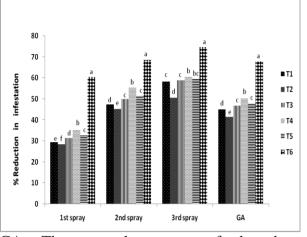


GA: The general average of the three successive sprays.

Bollworm infestation in Foliar application of *P. fluorescens* and *Beauveria basianna* treatment was significantly higher than that in the Profenophos 50 EC treatment, however, was significantly lower compared to the other treatments and the control. Reduction percentage in bollworms infestation was

Efficacy of *P. fluorescens* against pink bollworm

calculated and used to compare the bioinoculants and pesticide treatments (Fig. 2). **Fig. 2**. Percentages of the reduction bollworms infestation after applying *P. fluorescens* treatments in three successive sprays with 15 days interval. ANOVA, Tukey's at 0.05 level of probability; same alphabets during a counting indicates insignificant.



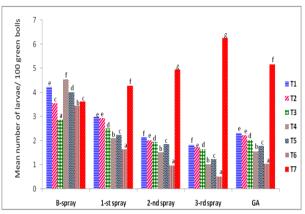
GA: The general average of the three successive sprays.

Reduction in the general average of infestation after the three sprays revealed that, there is no significant difference among the soil and foliar application of *P. fluorescens* and foliar application of *B. basianna* treatments. The percentage of reduction ranged between 41.4 % in Soil application of *P. fluorescens* 2.5 kg/ha treatment and up to 67.89 % in Profenophos 50 EC @ 1 lit/ha.

The foliar application of *P. fluorescens* and *B.* basianna treatment, exhibited the highest reduction the infestation that in was significantly different compared to the profenophos pesticide treatment. In general, mean number of larvae before and after spray was much lower compared to the level of infestation. Before spray, mean number of larvae ranged between 2.8 to 4.5 (Fig 3.). After three successive sprays with 15 days interval, mean number of larvae averaged 1% in pesticide treatment compared to 1.5% in foliar application of P. fluorescens @1% & B. basianna @ 1%, treatment and 5.16 % in the control (Fig 3.). As shown in Fig. 4, the percentage of reduction in the general average of larvae counted in profenophos pesticide

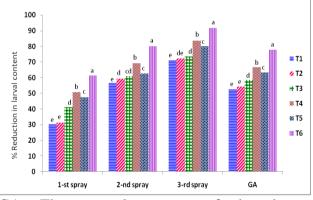
treatment was significantly higher compared to other treatments.

Fig. 3. Mean number of bollworms larvae before and after applying *P. fluorescens* treatments in three successive sprays with 15 days interval. ANOVA, Tukey's at 0.05 level of probability; same alphabets during a counting indicates insignificant



GA: The general average of the three successive sprays.

Fig. 4. Percentage of the reduction in bollworms larval content after applying *P*. *fluorescens* treatments in three successive sprays with 15 days interval. ANOVA, Tukey's at 0.05 level of probability; same alphabets during a counting indicates insignificant



GA: The general average of the three successive sprays.

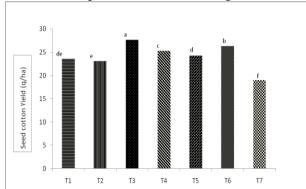
The above treatments exhibited 53 to 78 % reduction in larval content. The treatment of profenophos resulted in 77.78% reduction in the larval content. The other treatments could be arranged descendingly as follows: foliar application of *P. fluorescens* and *B. basianna*,

27

foliar application of B. basianna, soil and foliar application of *P. fluorescens*, soil application of P. fluorescens 2.5 kg/ha, foliar application of *P. fluorescens*.

The highest seed cotton yield was obtained from soil and foliar application of *P*. fluorescens followed by the profenophos pesticide treatment. The difference in seed cotton yield was significant in the treatments of bioinaculants. Control treatment had the lowest seed cotton yield that was significantly less compared to all other treatments.

Fig. 5. Cotton seed yield in control and P. fluorescens treatments. ANOVA. Tukev's Test, same alphabets indicates insignificant



DISCUSSION

Plant growth-promoting rhizobacteria (PGPR) which inhibiting in the rhizosphere, around/on the root surface, which improve the plant growth directly or indirectly. Qureshi and Ahmed (1991) proposed that spiny bollworm an economic injury when caused the infestation level reached 10%. However, Sing and Sandy (1993) and Purohit and Deshoande (1994) suggested that the chemical application should be done at 5% level of infestation. Saravanakumar et al. (2008), reported that application of fluorescent Pseudomonasds either individually or in combinations significantly reduced the leaf folder attack in rice. Commare et al. (2002) and Karthiba et al. (2010), revealed that the P. fluorescens strains was demonstrated to simultaneously reduce the incidence of a herbivorous insect, the rice leaf folder and a phytopathogenic fungus in rice under greenhouse and field conditions. Raiendran et al. (2007)demonstrated the PGPR and endophytic bacteria mediated induction of defense

28

responses in cotton plants against bollworm (Helicopverpa armigera) insect pest. In addition Pseudomonas rhizobacteria have been reported to stimulate plant growth under different condition in vitro (Saravanakumar and Samiyappan, 2007). Similar to this Williams and Asher (1996), demonstrated that Pseudomonas spp. significantly improved the emergence and proportion of healthy seedlings in sugar beet when compared with untreated seeds. Further, Vivekananthan et al. (2004) reported that application of *P. fluorescens* increased the fruit yield in mango. Similar to previous findings, the current study also documented the reduced bollworm infestation and increased the seed cotton yield in plots treated with P. fluorescens. Our study revealed that the bio inoculants had a significant influence on bollworms reduction and high yield in cotton and can be best utilized for cotton IPM programmes. Further, PGPR showed more consistent improvements of cotton, highlighting the importance of inoculant selection as reported by Araújo et al. (2018).

ACKNOWLEDGEMENT

The authors acknowledge the Vanavarayar Institute of Agriculture and The Southern India Mills' Association (SIMA) for providing support for conducting the experiments Thanks to Tropical Agro Pvt successfully. Ltd., Chennai for providing the bio inoculants for the study.

REFERENCE

- Araújo, G. C., Sousa, N. R., Ramos, M. A., Vega, A. L. and Castro, P. M. 2018. Performance of Quercus suber L. at nursery stage-application of two bio-inoculants under two distinct environments. Annals of Forest Science, 75(1): 29.
- Commare, R. R., Nandakumar, R., Kandan, S., Bharathi, A., Suresh, M. and T. Raguchabdar, 2002. Pseudomonas fluorescens based bioformulation forth management of sheath blight and leaf folder insect in rice. Crop Protection, 21: 671-677.
- EI- Naggar, A. Z. 1998. Evaluation of certain foliars and microelements in an Integrated

Pest Management (IPM) program tp control cotton bollworms, MSc Thesis, Facalty of Agriculture, Alexandria University, E

- Gomez, A. K. and Gomez A. A. 1984. Statistical procedures for agricultural research. John Wiley and Sons. Inc., Singapore.
- Haque, H. 1991. Imported generic pesticides need to be checked before marketing.Pakistan Agriculture Pesticides Association, *Bulletin*, 6: 16-17.
- Henderson, C.F. and Tilton, E.W. 1955. Test with acaricide against the brown wheat mite. *Journal of Economic Entomology*, **48**: 157-161
- Hussein, N. M., Shalaby, F. F., EI- Khayat,
 E.F., Tawfik, S.M. and Salem, M. S. 2002.
 Effect of certain agrochemicals on cotton a growth and bollworms infestation during three successive seasons at Kalub,
 Governorate, Egypt Proceeding of the 2nd International conference. Plant Protection Research Institute. December 21-24. 2002 Cario Egypt, **PP** 845-865.
- Karthiba, I., Saveetha, K., Suresh, S., Raguchander, I., Saravanakumar, D. and Samiyappan, R. 2010. PGPR and endomopathogenic fungus bioformulation for the synchronous management of leaf folder pest and sheath blight disease of rice. *Pest Manegement Sciences.* **66:** 555-564.
- Purohit, M. S. and Deshonade, A. D. 1994. Effects of fertilizer on Cotton bollworms in relation to plant protection. *Journal* of Maharashtra Agricultural Universities, 19: 172-174.
- Qureshi, Z. A. and Ahmed, N. 1991. Monitoring seasonal population fluctuation of spotted and spiny bollworm by synthetic sex pheromones and its relationship to boll infestation in cotton. *Journal of Applied Entomology*, **112**: 171-175.
- Rajendran, L. Samiyappan, R. Raguchander, T. and Saravanakumar, D. 2007.Endophytic bacteria mediate plant resistance against cotton bollworm. *Journal of Plant Interactions*, 2: 1-10.

Saravanakumar, D. and Samiyappan, R. 2007. ACC deaminase from *Pseudomonas fluorescens* mediated saline resistance in groundnut plants. *Journal of Applied Microbiology*, **102:** 1283-1292.

- Saravanakumar, D., Lavanya, N., Muthumeena, B., Raguchaner, T., Suresh, S. and Samiyappan, R. 2008. *Pseudomonas fluorescens* enhances resistance and natural enemy population in rice plants against leaf folder pest. *Journal of Applied Entomology*, **132:** 469-479.
- Sing, J. and Sandy, B. S. 1993. Economic threshold for bollworms control on arboretum cotton, *Cotton Research And Development Association*, **7:** 285-288.
- Vivekananthan, R., Ravi, M., Ramanathan, A. and Samiyappan, R. 2004. Lytiv enzyme induced by *Pseudomonas fluorescens* and other biocontrol organisms mediate defense against the anthracnose pathogen in mango. *World Journal of Microbiology and Biotechnology*, **20:** 235 – 244.
- Williams, G. E. and Asher, M. J. C. 1996. Selection of rhizobacteria for the control of Phythiumultimum onsugarbeet seedlings. *Crop Protection*, **15:** 479-486.

Manjula, T.R.^{1*}, Kannan, G.S.² and Sivasubramanian, P.³

¹Vanavarayar Institute of Agriculture, Department of Entomology, Manakkadavu, Pollachi-103, India

²The Gandhigram Rural Institute - Deemed University, Faculty of Agriculture and Animal Husbandry, Gandhigram, Dindigul District, Tamil Nadu, India.

³Tamil Nadu Agriculture University, Department of Entomology, Coimbatore-641003, Tamil Nadu, India. ***Corresponding author**

E-mail: manjulatr@gmail.com

29