In vitro evaluation of antagonistic activity of *Pseudomonas fluorescens* against fungal pathogen

Manoj Kumar Maurya*, Ramji Singh and Ajay Tomer

ABSTRACT

The present investigation was undertaken to isolate different strains of *Pseudomonas fluorescens* from various agroecological zones or crop's rhizosphere like moong, brinjal, rice, chilli, mustard, chirchida and tomato. Totally eight micro flora resembling *Pseudomonas fluorescens* were isolated and three isolates were confirmed as *P. fluorescens* (strain P.f.01, strain P.f.05 and strain P.f.07). *Pseudomonas fluorescens* strains P.f 07 were found most effective with the highest antagonistic activity against three fungal pathogen and show maximum inhibition of mycelial growth of *Fusarium moniliforme* (65.45%), *Rhizoctonia solani* (68.23%), and *Alternaria alternate* (48.13%).

MS History: 6.2.2014 (Received)-10.4.2014 (Revised)-14.6.2014 (Accepted)

Key words: Antagonistic activity, biocontrol agent, isolation, Pseudomonas fluorescens

INTRODUCTION

Pseudomonas fluorescens encompasses a group of common, gram negative, rod shaped, non pathogenic saprophytes bacteria that colonize in soil, water and plant surface environments. Since they are well adapted in soil, P. fluorescens strains are being investigated extensively for use in biocontrol of pathogens in agriculture (Ganeshan and Kumar, 2006). Biological control of plant pathogens by antagonistic micro organisms is a potential non-chemical means (Harman, 1991) and is known to be a cheap and effective eco-friendly method for the management of crop diseases (Cook and Baker, 1983). Pseudomonas fluorescens is adapted to survival in soil and colonization of plant roots (Kiely et al., 2006) and this applies also to the particular case of biocontrol agents from this species. Biocontrol strains have noticeably been observed at the root surface, (i.e. the rhizoplane) often forming microcolonies or discontinued biofilms in the grooves between epidermal cells. Certain strains are also capable of endophytic colonization. Within root tissues, they are mostly found in the intercellular spaces of the epidermis and the cortex (Duijff et al., 1997). Many biocontrol agents from P. fluorescens and closely related species are well characterized for their ability to produce antimicrobial compounds, including 2,4diacetylphloroglucinol (DAPG), phenazines, hydrogen cyanide and surfactants (Haas and De'fago, 2005). The use of biological control agents as an alternative to fungicides is increasing rapidly in the present day agriculture, due to the deleterious effects of chemical pesticides on human health and environment.

MATERIALS AND METHODS

Present investigations on *Pseudomonas fluorescens*, an antagonistic bacterium with special reference to evaluation of antagonistic activity of *P. fluorescens* against fungal pathogen were carried out in the laboratory of the Department of Plant Pathology of Sardar Vallabhbhai Patel University of Agriculture and Technology, Meerut (U.P.).

Isolation

For isolation, one gram of soil sample was placed in a 250 mL conical flask containing 100 mL of sterilized distilled water (SDW) and mixed thoroughly. Different dilutions of working samples were prepared by serially diluting the stock solution upto 10^{-8} . One ml of last serial dilution *i.e.*, 10^{-8} was spread on *P. fluorescens* selective King's B Medium (King's *et al.*, 1954) for isolation of *P. fluorescens*. The plates were incubated for 2 days at $28\pm2^{\circ}$ C and after incubation, pure culture was grown; colour of bacterial colony was initially yellow but turned yellow green as pigmentation were produced (Bonds, 1957).

Isolation of *Pseudomonas fluorescens* from different rhizosphere

To isolate the *P. fluorescens* from different rhizosphere, the soil samples were collected from different location as given below:

Isolates No.	Vegetation	Location	Tentative Identification
P.f.01	Moong	PCP SVP UNI. Meerut	P.fluorescens
P.f.03	Brinjal	HRC SVP UNI. Meerut	P.fluorescens
P.f.05	Rice	CRC Chiraudi SVP UNI.Meerut	P.fluorescens
P.f.07	Chilli	Farmer field Meerut	P.fluorescens
P.f.09	Chilli	PCP SVP UNI. Meerut	P.fluorescens
P.f.11	Mustard	Farmer field Meerut	P.fluorescens
P.f.12	Chirchida	Meerut	P.fluorescens
P.f.13	Tomato	Farmer field Ambedkar Nagar	P.fluorescens

Culture and maintenance

Medium consists of peptone 20gm, agar-agar 20gm, monophosphate 1.5gm. (K_2HPO_4) potassium magnesium sulphate (MgSo4) 1.5gm, glycerol 10mL and distilled water 1000 mL (King et al., 1954). After mixing all the ingredients with distilled water, media was placed into a stainless steel pan and stirred with glass rod for proper mixing of all the ingredients. Now the medium was filtered through a muslin cloth by squeezing out whole liquid. 200 mL medium was placed in each 500 mL capacity flasks. Flasks were tightly plugged with non-absorbent cotton plug and wrapped with butter paper and rubber band. Medium was autoclaved at 15 psi (121.6°C) for 20 min and cooled before pouring into Petri plates.

Another medium contain agar–agar 20.0g, dextrose 20.0g, potato (peeled and sliced) 200.0g, distilled water 1 L. 250g of potato was peeled and cut into small and fine sliced pieces. Exactly 200g of potato pieces were weighed and placed into a stainless steel pan. 500 mL of water was added to potato pieces and boiled gently for such a period until they are easily mashed by a glass rod. The decoction was filtered through muslin cloth and squeezed out all the liquid in a measuring cylinder and potato pieces were discarded. Now sufficient amount of water was added to make the volume 1000 mL. Now preweighed agar agar was added (20g) bit by bit to the boiling solution to dissolve it. At the same time

dextrose (20g) was also added in boiling solution (melted with agar) and final volume made up to 1 L. It was poured @ about 200 mL in each of four conical flasks of 500mL 2ycelia2 and 10 mL per culture tube to prepare the PDA slants. Both, flasks and culture tubes were tightly plugged with nonabsorbent cotton and wrapped with butter paper and rubber bands. The culture tubes and flasks were placed vertically (mouths up) in wire baskets and then autoclaved at 15 psi (121.6°C) for 20 min. The bacteria, initially isolated in a pure culture on King's B media and sub cultured on PDA slants were allowed to grow at $28\pm2^{\circ}C$ temperature. The culture thus obtained was stored in refrigerator at 5^{0} C for further studies and was sub cultured periodically.

Antagonism of *P. fluorescens*

The antagonistic activity of *P. Fluorescens* against Fusarium moniliforme, Rhizoctonia solani and Alternaria alternata were tested by dual culture technique. Bacterial isolates were streaked at one side of Petri dish (one cm away from the edge) containing PDA. 9 mm mycelial disc from seven days old PDA culture of F. moniliforme, R. solani, and A. alternata were placed at the opposite side of Petri dishes perpendicular to the bacterial streak respectively and incubated at $27\pm2^{\circ}$ C for 5-7 days. Petri dishes inoculated with fungal discs alone replications control. Three served as were maintained for each isolate. Observations on width of inhibition zone and mycelial growth of test pathogen were recorded and per cent inhibition of pathogen growth was calculated by using the formula proposed by Vincent (1927).

Per cent inhibition (I) = C-T/C $\times 100$

Where.

C- mycelial growth of pathogen in control

T- mycelial growth of pathogen in dual culture plate.

RESULTS AND DISCUSSION

Isolation of P. fluorescens

Eight microflora resembling *P. fluorescens* were isolated from the rhizhosphere of different crops/plants.On the basis of colony characters and pigmentation etc, out of eight microflora, three isolates were confirmed as *P. fluorescens* and tested against *Fusarium moniliforme, Rhizoctonia solani,* and *Alternaria alternata*. The three *Pseudomonas*

Antagonistic activity of Pseudomonas fluorescens

fluorescens isolates isolated and confirmed as *Pseudomonas fluorescens* during present investigation were designated as given below:

P.f.	Source	Location
Isolates	(Rhizosphere)	
P.f.01	Moong	P.C.P. SVPUAT, Meerut.
P.f.05	Rice	C.R.C.
		Chiraudi,SVPUAT,Meerut.
P.f.07	Chilli	Vill. Siwaya Jamalullahpur,
		Meerut.

Dual culture technique

The results of the dual culture technique indicated that the three isolates inhibited growth of tested fungi significantly. In case of *Fusarium moniliforme*, a maximum inhibition of 65.45% was recorded by PF 07 and minimum of 45.45% was recorded with the isolate P.f.05. In case of *Rhizoctonia solani*, the maximum inhibition of 68.23% was exhibited by PF 07 and minimum, 55.8% was recorded with the P.f.05 isolate, where as in case of *Alternaria alternata* maximum inhibition 48.13% was recorded by by P.f.07 and minimum, 44.40% was recorded with isolate P.f.05 (Table 1).

Table 1. Antagonistic activity of *Pseudomonas fluorescens*against chosen fungi

P.f. Isolates	Fungi	Mycelial growth (mm)	Growth inhibition (%)
P.f.01	F. moniliforme	11.00	60.00
P.f.05		15.00	45.45
P.f.07		9.50	65.45
Control		27.50	0
CD @ 5 % =			5.139
P.f.01	R. solani	34.50	59.41
P.f.05		37.50	55.88
P.f.07		27.00	68.23
Control		85.00	0
	CD @ 5% =		12.011
P.f.01	A. alternata	49.00	45.14
P.f.05		49.66	44.40
P.f.07		46.33	48.13
Control		89.33	0
	CD @ 5 % =		5.736

Attempst were made to isolate and maintain different strains of *Pseudomonas fluorescens* from different agroecological zones, so that they can be used in future for stress management in various crops in varying agroecological situations. In this direction eight microflora were isolated on King's B medium and further they were subjected to identification on the basis of antagonistic characteristics exhibited against some pathogenic microflora. Out of eight microflora isolated on King's B medium, only three were confirmed as *P. fluorescens* (isolate P.f 01, isolate P.f 05 and isolate P.f 07), which neither showed fluorescent pigment production nor they were able to inhibit any of the fungal pathogen when subjected to dual culture. The isolate P.f 01 was isolated from the rhizosphere of Moong (*Vigna radiata*) from the field of P.C.P. SVPUAT, Meerut. The isolate P.f.05-was isolated from the rhizosphere of rice crop from the field C.R.C. Chiraudi of SVPUAT, Meerut. The isolate P.f.07-was isolated from the rhizosphere of chilli crop from a farmer field of village Siwaya Jamalullahpur district of Meerut.

The results of the dual culture technique indicated that the three isolates were able to inhibit growth of tested fungal pathogen significantly. In case of F. moniliforme a maximum and minimum inhibition was exhibited by P.f 07 and P.f 05 respectively. In case of Rhizoctonia solani, the maximum inhibition of 68.23% was exhibited by P.f 07 and minimum, 55.8% was exhibited by P.f 05. where as in case of Alternaria alternata maximum inhibition 48.13% was exhibited by P.f 07 and minimum, 44.40% was exhibited with isolate PF05. Vanitha et al. (2007) screened six isolates of Pseudomonas fluorescens against Alternaria chlamydospora Mouchacca causing leaf blight diseases in Solanum nigrum L under in vitro conditions. The experimental results showed that Pf MMP of Pseudomonas showed 100 per cent inhibition of mycelial growth and culture filtrate of P. fluorescens (PfMMP) significantly reduced the mycelial growth and resulted in minimum spore germination of A. Chlamydospora. Adhikari et al. (2013) reported that among seventy isolates, antagonistic twenty one representing biovars of P. fluorescens (biovars I, II, III, and V) were collected from the rhizosphere of okra, chilli, ground nut, brinjal, cabbage and tomato from different agroecological regions of West Bengal and were subjected to evaluation for their antifungal activity under in vitro condition against Rhizoctonia solani, the most important soil-borne plant pathogen. Two isolates of P. fluorescens PF-8 and PF-7 effectively inhibited the mycelial growth of Rhizoctonia solani in dual culture method. The vigour index of okra was also recorded maximum for the isolate P.f 08 than by P.f 07.

Different strains of *P. fluorescens* from different agro-ecological zones, so that they can be used in future for stress management in various crops in

Manoj Kumar Maurya et al.

varying environment. The isolates confirmed as *P. fluorescens* were subjected to testing of antagonistic activity of *Pseudomonas fluorescens* against three fungal pathogen. Among three tested isolates of *Pseudomonas fluorescens*, strain P.f 07 was exhibited maximum antagonistic activity against fungal pathogen, *Fusarium moniliforme, Rhizoctonia solani* and *Alternaria alternata*.

REFERENCES

- Adhikari, A. Dutta, S. Nandi, S. Bhattacharya, I. Roy, M. de Sarkar, G. Mandal, T. 2013. Antagonistic potentiality of native rhizobacterial isolates against root rot disease of okra, incited by *Rhizoctonia solani*, *African Journal of Agricultural Research*, 8 (4): 405-412.
- Bonds, G.J., Jensen, C.E., and Thamasen, J., 1957. A water soluble fluorescing bacterial pigment which deplomerize hylorenic acid. *Acta Pharmocol*, *Toxicol*, **13**: 184-193.
- Cook, R.S. and Baker, K.F.1983, The nature and practice of biological control of plant pathogens. *American Phytopathological Society, St Paul, Minn*, 539 **PP**.
- Duijff, B.J., Gianinazzi-Pearson, V. and Lemanceau, P. 1997. Involvement of the outer membrane lipopolysaccharides in the endophytic colonization of tomato roots by biocontrol Pseudomonas fluorescens strain WCS417r. *New Phytologist*, **135**: 325–334.
- Ganeshan, G. and Kumar, M.A. 2006. Pseudomonas fluorescens, a potential bacterial antagonist to control plant diseases. Journal of Plant Interactions, 1(3): 123-134.

- Haas, D. and De'fago, G. 2005. Biological control of soilborne pathogens by fluorescent pseudomonads. *Nature Reviews Microbiology*, **3**: 307–319.
- Harman, G.E. 1991. Seed treatment for biological control of plant diseases, *Crop Protection*, **10**: 166-171.
- Kiely, P.D., Haynes, J.M., Higgins, C.H., Franks, A., Mark, G.L., Morrissey, J.P. and O'Gara, F. 2006. Exploiting new systems-based strategies to elucidate plant-bacterial interactions in the rhizosphere. *Microbial Ecology*, **51**: 257–266.
- King, E.O., Ward, M.K. and Raney, D.E. 1954. Two simple media for the demonstration of Pyocyanin and fluorescin. *Journal of Laboratory and Clinical Medicine*, **36**: 100-102.
- Vanitha, S. and Samiyappan, R. 2007. Screening of bacterial antagonistic micro-organisms under in vitro conditions against Alternaria chlamydospora causing leaf blight disease in Solanum nigrum L. *Biomed*, 2(2): 155-163.

Manoj Kumar Maurya*, Ramji Singh and Ajay Tomer

Department of Plant Pathology, Sardar Vallabhbhi Patel University of Agriculture and Technolody, Meerut- 250110 (UP), India.

*E-mail: manoj.maurya0805@gmail.com